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(54) **Title:** NOVEL A-GALACTOSIDASES

(57) **Abstract:** This invention relates to novel α -galactosidases for the enzymatic removal of the immunodominant monosaccharides on blood products and tissues. Specifically this invention provides a novel family of $\alpha 3$ glycosidases, used for the enzymatic removal of type B antigens from blood group B and AB reactive blood products, and the GalIIi antigen from non-human animal tissues, thereby converting these to non-immunogenic cells and tissues suitable for transplantation.

NOVEL α -GALACTOSIDASES

FIELD OF THE INVENTION

This invention relates to a novel family of polypeptides having α -galactosidase activities, demonstrating unique substrate specificities and superior kinetic properties, that are used for
5 removal of the immunodominant monosaccharides on blood products and tissues. Specifically this invention provides a novel family of α 3 glycosidases, used for the enzymatic removal of type B antigens from blood group B and AB reactive blood products, and the Galili antigen from non-human animal tissues, thereby converting these to non-immunogenic cells and tissues suitable for transplantation.

10 BACKGROUND OF THE INVENTION

As used herein, the term "blood products" includes whole blood and cellular components derived from blood, including erythrocytes (red blood cells) and platelets.

There are more than thirty blood group (or type) systems, one of the most important of which is the ABO system. This system is based on the presence or absence of antigens A and/or
15 B. These antigens are found on the surface of erythrocytes and platelets as well as on the surface of endothelial and most epithelial cells. The major blood product used for transfusion is erythrocytes, which are red blood cells containing hemoglobin, the principal function of which is the transport of oxygen. Blood of group A contains antigen A on its erythrocytes. Similarly, blood of group B contains antigen B on its erythrocytes. Blood of group AB contains both antigens, and
20 blood of group O contains neither antigen.

The blood group structures are glycoproteins or glycolipids and considerable work has been done to identify the specific structures making up the A and B determinants or antigens. The ABH blood group specificity is determined by the nature and linkage of monosaccharides at the ends of the carbohydrate chains. The carbohydrate chains are attached to a peptide
25 (glycoprotein) or lipid (glycosphingolipid) backbone, which are attached to the cell membrane of the cells. The immunodominant monosaccharide determining type A specificity is a terminal α 1-3 linked *N*-acetylgalactosamine (GalNAc), while the corresponding monosaccharide of B type specificity is an α 1-3 linked galactose (Gal). Type O cells lack either of these monosaccharides at the termini of oligosaccharide chains, which instead are terminated with α 1-2 linked fucose (Fuc)
30 residues.

A great diversity of blood group ABH carbohydrate structures are found due to structural variations in the oligosaccharide chains that carry ABH immunodominant saccharides. Table 1 lists structures reported in man and those that have been found on human red cells or in blood
35 extracts. For a review, see, Clausen & Hakomori, *Vox Sang* 56(1): 1-20, 1989). Red cells contain ABH antigens on N-linked glycoproteins and glycosphingolipids, while it is generally believed that

O-linked glycans on erythrocytes glycoproteins, mainly glycophorins, are terminated by sialic acid and not with ABH antigens. Type 1 chain glycosphingolipids are not endogenous products of red cells, but rather adsorbed from plasma.

5 **Table I:**
Histo-Blood Group ABH Immunoreactive Determinants of Human Cells¹

Name	Hapten Structure	Type of Glycoconjugate	Found on RBC	No
A type 1, ALe ^d	GalNAc α 1-3Gal β 1-3GlcNAc β 1-R 2 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid	1
A type 1, ALe ^b	GalNAc α 1-3Gal β 1-3GlcNAc β 1-R 2 4 Fuc α 1 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid	2
A type 2, A	GalNAc α 1-3Gal β 1-4GlcNAc β 1-R 2 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid N-linked	3
A type 2, ALe ^y	GalNAc α 1-3Gal β 1-4GlcNAc β 1-R 2 3 Fuc α 1 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid?	4
A type 3, O-linked	GalNAc α 1-3Gal β 1-3GalNAc α 1-O-Ser/Thr 2 Fuc α 1	O-linked		5
A type 3, Repetitive	GalNAc α 1-3Gal β 1-3GalNAc α 1-3Gal β 1-4GlcNAc β 1-R 2 2 Fuc α 1 Fuc α 1	Glycolipid	Glycolipid	6
A type 4, Globo	GalNAc α 1-3Gal β 1-3GalNAc β 1-3Gal α 1-R 2 Fuc α 1	Glycolipid	Glycolipid?	7
A type 4, Ganglio	GalNAc α 1-3Gal β 1-3GalNAc β 1-3Gal β 1-R 2 Fuc α 1	Glycolipid		8
B type 1, BLe ^d	Gal α 1-3Gal β 1-3GlcNAc β 1-R 2 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid	9
B type 1, BLe ^b	Gal α 1-3Gal β 1-3GlcNAc β 1-R 2 4 Fuc α 1 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid	10
B type 2, B	Gal α 1-3Gal β 1-4GlcNAc β 1-R 2 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid N-linked	11
B type 2, BLe ^y	Gal α 1-3Gal β 1-4GlcNAc β 1-R 2 3 Fuc α 1 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid?	12
B type 3, O-linked	Gal α 1-3Gal β 1-3GalNAc α 1-O-Ser/Thr 2 Fuc α 1	O-linked		13
B type 4, Globo	Gal α 1-3Gal β 1-3GalNAc β 1-3Gal α 1-R 2 Fuc α 1	Glycolipid?	Glycolipid?	14
B type 4, Ganglio	Gal α 1-3Gal β 1-3GalNAc β 1-3Gal β 1-R 2 Fuc α 1	Glycolipid?		15
H type 1, Le ^a	Gal β 1-3GlcNAc β 1-R 2 Fuc α 1	Glycolipid N-linked O-linked	Glycolipid	16

Name	Hapten Structure	Type of Glycoconjugate	Found on RBC	No
H type 1, Le ^b	Galβ1-3GlcNAcβ1-R 2 4 Fucα1 Fucα1	Glycolipid N-linked O-linked	Glycolipid	17
H type 2, H	Galβ1-4GlcNAcβ1-R 2 Fucα1	Glycolipid N-linked O-linked	Glycolipid N-linked	18
H type 2, Le ^y	Galβ1-4GlcNAcβ1-R 2 3 Fucα1 Fucα1	Glycolipid N-linked O-linked	Glycolipid?	19
H type 3, O-linked	Galβ1-3GalNAcα1-O-Ser/Thr 2 Fucα1	O-linked		20
H type 3, H-A	Galβ1-3GalNAcα1-3Galβ1-4GlcNAcβ1-R 2 2 Fucα1 Fucα1	Glycolipid	Glycolipid (A RBC)	21
H type 4, Globo	Galβ1-3GalNAcβ1-3Galα1-R 2 Fucα1	Glycolipid	Glycolipid	22
H type 4, Ganglio	Galβ1-3GalNAcβ1-3Galβ1-R 2 Fucα1	Glycolipid		23
Thomsen-Friedenrich Tf, T	Galβ1-3GalNAcα1-O-Ser/Thr	O-linked	O-linked (+SA)	24
Gal-A, T cross-react.	Galβ1-3GalNAcα1-3Galβ1-4GlcNAcβ1-R 2 Fucα1	Glycolipid	Glycolipid (A RBC)	25
Tn, A cross-react.	GalNAcα1-O-Ser/Thr	O-linked	O-linked (+SA)	26

¹Adapted from Clausen and Hakomori, *Vox Sang* 56(1): 1-20, 1989. Designations: "?" indicates potential glycolipid structures which have not been reported to date.

- Blood group A and B exist in several subtypes. Blood group A subtypes are the most frequent, and there are three recognized major sub-types of blood type A. These sub-types are
- 5 known as A₁, A intermediate (A_{int}) and A₂. There are both quantitative and qualitative differences that distinguish these three sub-types. Quantitatively, A₁ erythrocytes have more antigenic A sites, *i.e.*, terminal *N*-acetylgalactosamine residues, than A_{int} erythrocytes which in turn have more antigenic A sites than A₂ erythrocytes. Qualitatively, A₁ erythrocytes have a dual repeated A structure on a subset of glycosphingolipids, while A₂ cells have an H structure on an internal A
- 10 structure on a similar subset of glycolipids (Clausen *et al.*, *Proc. Natl. Acad. Sci. USA* 82(4): 1199-203, 1985, Clausen *et al.*, *J. Biol. Chem.* 261(3): 1380-7, 1986). These differences between A₁ and weak A subtypes are thought to relate to differences in the kinetic properties of blood group A isoenzyme variants responsible for the formation of A antigens (Clausen *et al.*, *J. Biol. Chem.* 261(3): 1388-92, 1986). The differences of group B subtypes are believed to be solely of
- 15 quantitative nature.

Blood of group A contains antibodies to antigen B. Conversely, blood of group B contains antibodies to antigen A. Blood of group AB has neither antibody, and blood group O has both.

Antibodies to these and other carbohydrate defined blood group antigens are believed to be elicited by continuous exposure to microbial organism carrying related carbohydrate structures. An individual whose blood contains either (or both) of the anti-A or anti-B antibodies cannot receive a transfusion of blood containing the corresponding incompatible antigen(s). If an individual receives a transfusion of blood of an incompatible group, the blood transfusion recipient's antibodies coat the red blood cells of the transfused incompatible group and cause the transfused red blood cells to agglutinate, or stick together. Transfusion reactions and/or hemolysis (the destruction of red blood cells) may result therefrom.

In order to avoid severe transfusion reactions due to the presence of antibodies to the A and B blood group antigens the blood group of the donor and the recipient are matched before blood transfusions by typing methods. For example, a blood type A recipient can be safely transfused with type A blood, which contains compatible antigens, but not type B blood, which would trigger an adverse immune response in the recipient. Because type O blood contains no A or B antigens, it can be transfused into any recipient with any blood type, *i.e.*, recipients with blood types A, B, AB or O. Thus, type O blood is considered "universal", and may be used for all transfusions. Hence, it is desirable for blood banks to maintain large quantities of type O blood. However, there is a paucity of blood type O donors. Therefore, it is desirable and useful to remove the immunodominant A and B antigens on types A, B and AB blood in order to maintain large quantities of universal blood products.

In an attempt to increase the supply of type O blood, methods have been developed for converting type A, B and AB blood to type O blood. Although, enzymatic conversion of both group B and group A red cells have been achieved in the past, these older processes have several disadvantages, particularly that they require excessive quantities of enzyme, and the specificities of many glycan modifying enzymes are not restricted to cleavage of only the blood group A or B antigens.

As will be explained below, the present invention provides for a family of polypeptides having highly refined substrate specificities, and better kinetic properties, that can be used to generate tissues and blood products lacking immunodominant antigens, thereby providing an efficient and cost-effective commercial process to supply, *e.g.* universal (non-immunogenic) blood cells for transplant, and even animal tissues for xenotransplantation into humans.

CONVERSION OF BLOOD GROUP B CELLS:

Enzymatic conversion of type B blood using purified or recombinant Coffee bean (*Coffea canephora*) α -galactosidase has been achieved using 100-200 U/ml (U.S. Pat. No. 4,427,777; Zhu *et al.*, Arch Biochem Biophys 1996; 327(2): 324-9; Kruskall *et al.*, Transfusion 2000; 40(11): 1290-8). The specific activity of Coffee bean α -galactosidase was reported to be 32 U/mg using p-nitrophenyl α -D-Gal with one unit (U) defined as one μ mole substrate hydrolyzed per minute

(Zhu *et al.*, Arch Biochem Biophys 1996; 327(2): 324-9). Enzymatic conversions were done at pH 5.5 with approximately 6 mg/ml enzyme at 80-90% hematocrit, and the resulting converted O cells functioned normally in transfusion experiments and no significant adverse clinical parameters were observed (Kruskall *et al.*, Transfusion 2000; 40(11): 1290-8). This data along with earlier
5 publications, clearly demonstrate that enzymatic conversion of red blood cells is feasible and that such enzyme group B converted O (B-ECO) cells can function as well as matched type untreated cells in transfusion medicine. Nevertheless, the quantities of enzymes required for seroconversion in these studies, even with recombinant production of the enzyme, renders this method for generating ECO cells impractical mainly for economical reasons.

10 Claims of protocols for improved conversion of B cells using recombinant Glycine max α -galactosidase with a specific activity of approximately 200 U/mg have been reported using 5-10 units of enzyme/ml blood (with 16% hematocrit) (see, U.S. Pat. Nos. 5,606,042; 5,633,130; 5,731,426; 6,184,017). The Glycine max α -galactosidase was thus used at 25-50 μ g/ml, which represents a significant reduction in enzyme protein quantities required (50-200 fold) (Davis *et al.*,
15 Biochemistry and Molecular Biology International, 39(3): 471-485, 1996). This reduction is partly due to the higher specific activity of the Glycine max α -galactosidase (approximately 6 fold) as well as different methods used for conversion and evaluation. The 200 U/ml enzyme used in the study of Kruskall *et al.*, (Transfusion, 40(11): 1290-8, 2000) was worked out for full unit (approximately 220 ml packed cells) conversions at 80-90% hematocrits and thoroughly analyzed by standard
20 blood bank typing as well as by more sensitive cross-match analysis. Furthermore, the efficiency of conversion was evaluated by analysis of survival and induced immunity in patients receiving multiple transfusions of converted cells. The enzymatic conversions were done in test tubes in ml scale at 16% hematocrit, as described in U.S. Pat. No. 5,606,042 (and 5,633,130; 5,731,426; 6,184,017) with Glycine max α -galactosidase, and the conversion efficiency not evaluated by
25 cross-match analysis. Conversion of cells at 16% hematocrit required 10 U/ml, while conversions at 8% required 5 U/ml, indicating that converting at increased hematocrit requires more enzyme although higher cell concentrations were not tested. Thus, part of the reduction in enzyme protein quantities required compared to protocols reported by Kruskall *et al.*, (Transfusion 2000; 40(11): 1290-8), is related to the concentration (hematocrit) of cells used in conversion, and this may
30 represent more than 5-10 fold, although direct comparison is not possible without further experimentation. The U.S. Pat. No. 5,606,042 (and 5,633,130; 5,731,426; 6,184,017) further provides improvements in the conversion buffer using Na citrate and glycine at less acidic pH (preferably pH 5.8) and including additional protein in the form of BSA (bovine serum albumin) for stabilization. Interestingly, the conversion buffer developed for the Glycine max α -galactosidase
35 was found not to be applicable to Coffee bean α -galactosidase. Although, some improvement in the conversion of B cells may be provided by U.S. Pat. No. 5,606,042 (and 5,633,130; 5,731,426; 6,184,017), it is clear that at least more than 0.5 mg of enzyme is required per ml packed type B red cells using the disclosed protocol. It is likely that considerable more enzyme than this is

required to obtain cells fully converted to O cells by the most sensitive typing procedures used in standard blood bank typing protocols. Furthermore, the protocol requires introduction of additional extraneous protein (BSA or human serum albumin) as well as exposing blood products to a significant acidic pH.

5 Bakunina *et al.* (Bakunina *et al.* Biochemistry (Moscow) 1998, p1420) has claimed the identification and isolation of a novel α -galactosidase from the marine bacterium *Pseudoalteromonas* spp. (KMM 701). The isolated enzyme preparation was purified to a specific activity of 9.8U/mg using the substrate pNP-Gal and had an apparent molecular weight by gel filtration of 195kD. The enzyme preparation efficiently cleaved the monosaccharide substrate
10 pNP-Gal with an apparent K_m for pNP-Gal of 0.29 mM as well as several unbranched disaccharides with terminal α -galactose including melibiose and Gal α 1-3Gal, and hence does not show high specificity for blood group B. This enzyme will therefore cleave unbranched oligosaccharides with terminal α -Gal such as the linear B structure as well as the P₁ antigen. The enzyme was reported to have a neutral pH optimum (*i.e.*, a pH optimum ranging from about 6.5 to
15 about 7.7) and to convert blood group B cells with 24 h incubation reaction time to cells typing as group O cells. However, details of the conversion procedure and enzyme consumption were not described, and the efficiency of conversion evaluated by standard typing procedures with licensed typing reagents remains to be tested. Purification to homogeneity, cloning and recombinant expression of the enzyme will likely be required to provide the quantities and quality of enzyme
20 protein required for enzymatic conversion of red cells.

We have disclosed (U.S.S.N. 10/251,271) the identification and partial characterization of a novel α -galactosidase activity with high specific activity and highly restricted substrate specificity for the blood group B antigen. The enzyme activity was identified by screening more than 2,400 bacterial and fungal isolates and found in only a few bacteria. The enzyme was partly purified
25 from cell lysates of *Streptomyces griseoplanus* strain #2357 (ATCC deposit No. PTA-4077) and partial amino acid sequence information was obtained.

It is evident from the above that further improvements in conversion of B cells is required in order to make this a practical and commercially applicable technology. Necessary improvements include obtaining more efficient and specific α -galactosidase enzymes, which allow
30 conversion to take place preferable at neutral pH and without extraneous protein added.

ASSAYS TO DETERMINE α GAL CLEAVING GLYCOSIDASE ACTIVITIES:

Past methods for searching, identification and characterization of exo-glycosidases have generally relied on the use of simple monosaccharide derivatives as substrates to identify
saccharide and potential linkage specificity. Derivatized monosaccharide, or rarely
35 oligosaccharide, substrates include without limitation p-nitrophenyl (pNP), benzyl (Bz), 4-methyl-umbrelliferyl (Umb), and 7-amino-4-methyl-coumarin (AMC). The use of such substrates provides

easy, fast, and inexpensive tools to identify glycosidase activities, and makes large scale screening of diverse sources of enzymes practically applicable. However, the kinetic properties and fine substrate specificities of glycosidase enzymes may not necessarily be reflected in assays with such simple structures. It is also possible that novel enzymes with high degree of specificity and/or selective efficiency for complex oligosaccharide and unique glycoconjugate structures exists, but that these may have been overlooked and remain unrecognized due to methods of analysis. Thus, in order to identify and select the optimal exo-glycosidase for a particular complex oligosaccharide or glycoconjugate structure it is preferable to use such complex structures in assays used for screening sources of enzymes. Furthermore, preferred assays used for screening include selection for preferable kinetic properties such as pH requirement and performance on substrates, e.g., attached to the membrane of cells.

In prior studies, all α -galactosidases (EC 3.2.1.22) and α -N-acetylgalactosaminidases (EC 3.2.1.49) used for removing the B and A antigens of blood cells had been identified and characterized using primarily p-nitrophenyl monosaccharide derivatives. Interestingly, most of these α -galactosidase and α -N-acetylgalactosaminidase enzymes used in past studies are evolutionary homologs as evidenced by significant DNA and amino acid sequence similarities. Thus, the human α -galactosidase and α -N-acetylgalactosaminidase are close homologs (Wang *et al.*, J Biol Chem, 265: 21859-66, 1990), and other enzymes previously used in blood cell conversion including the chicken liver α -N-acetylgalactosaminidase, fungal acremonium α -N-acetylgalactosaminidase, and bacterial α -galactosidases all exhibit significant sequence similarities. Sequence analysis of all known O-glycoside hydrolases have been grouped in 85 distinct families based on sequence analysis, and the above mentioned α -galactosidases and α -N-acetylgalactosaminidases are grouped in family 27 (http://afmb.cnrs-mrs.fr/~pedro/CAZY/ghf_32.html). These enzymes are characterized by having a retaining mechanism of catalysis and use aspartic acid as the catalytic nucleophile (Henrissat, Biochem Soc Trans, 26(2): 153-6, 1998; Rye & Withers, Curr Opin Chem Biol, 4(5): 573-80, 2000). The primary structure of a bacterial α -N-acetylgalactosaminidase from *Clostridium perfringens* was reported to be dissimilar and non-homologous to eukaryote α -N-acetylgalactosaminidases (Calcutt *et al.* FEMS Micro Lett 214:77-80, 2002), and is grouped in a distantly related glycosidase family 36, which also contains α -galactosidases and α -N-acetylgalactosaminidases (http://afmb.cnrs-mrs.fr/~pedro/CAZY/ghf_32.html). The catalytic mechanism of this group of enzymes is predicted to be similar to that of enzymes from family 27 because some sequence similarity exists between enzymes of the two families.

SUMMARY OF THE INVENTION

The present invention provides compositions and methods for the enzymatic removal of type B antigens from blood group B and AB reactive blood products. Specifically, this invention

provides compositions and methods for the specific enzymatic removal of the immunodominant monosaccharides specifying the blood group B antigens, namely α 1,3-D-galactose.

In one embodiment, this invention provides a novel family of homologous polypeptides, which have 20% or more overall amino acid sequence identity with SEQ NO. 2). These
5 polypeptides exhibit α -galactosidase activity across a neutral pH optimum, and demonstrate high substrate specificity for blood group B or Galilli antigen structures. and no or insignificant activity with α Gal or α GalNAc-pNP monosaccharide substrates. Certain members of this family show no or insignificant activity with linear α 1-3 linked Gal. Other polypeptides in this family will cleave linear structures, but show no or insignificant activity with P antigens and Gal α 1-4 linkages. The
10 novel polypeptides and gene family disclosed by the present invention has applications for use in the removal of immunodominant monosaccharides, α Gal and Galilli, from complex oligosaccharide targets such as those close to the true A and B carbohydrate antigens of the surface of cells in blood products, and the Galilli antigen from animal tissues.

In another aspect, this invention provides methods for the sero-conversion of all blood
15 group AB and B red cells, resulting in the removal of immunodominant B antigens from type B and AB cells. The removal of B antigens can be determined, for example by standard blood bank serological typing. According to the methods of this invention, the B antigens are removed using the polypeptides described, that (i) have highly restricted specificity for blood group B antigens, (ii) have optimal performance at neutral pH with blood group oligosaccharides; and (iii) are active
20 in red blood cell conversion at a slightly acidic to slightly basic, and preferably a neutral pH (pH from about 6 to about 8). These methods comprise the steps of: (a) contacting a blood product with one or more of these polypeptides, under approximately neutral pH conditions, for a period of time sufficient to remove the immunodominant B antigens, and (b) removing the polypeptide and digested antigen fragments from the seroconverted blood product.

In another embodiment, this invention provides methods for the removal of all detectable B
25 antigens from group B or AB red cells, using α -galactosidases that (i) have highly restricted specificity for blood group B antigens; and (ii) are active in red blood cell conversion with blood group oligosaccharides, over an approximately neutral pH range (pH about 6 to about 8).

In another aspect of the present invention, there are provided sero-converted erythrocytes.
30 In one embodiment, the sero-converted erythrocytes are characterized as: (i) having been converted from a type B or type AB erythrocyte to a non-B erythrocyte (having no detectable B antigens, as determined by standard blood bank serological typing) seroconverted using a polypeptide of the family described herein.

In yet another aspect, the invention includes a modified red blood cell comprising: a group
35 B erythrocyte or a group AB erythrocyte that lacks immunodominant B epitopes but displays α 1-4Gal epitopes, including the P1 and P^k blood group antigens. In one embodiment, the blood

cell substantially lacks immunodominant B epitopes as determined by serological typing or hemagglutination assays, and also lacks linear α 1-3 linked Gal structures. In another embodiment, the blood cell substantially lacks immunodominant B epitopes as determined by serological typing or hemagglutination assays, but retains linear α 1-3 linked Gal structures.

5 In yet another aspect, the invention includes a modified red blood cell prepared by the method comprising: obtaining a group B erythrocyte or a group AB erythrocyte, suspending the erythrocyte in a buffer solution having an approximately neutral pH (about pH 6 to about pH 8), and contacting the erythrocyte with an alpha galactosidase polypeptide thereby substantially cleaving from the erythrocyte the immunodominant B epitopes. In various embodiment, the
10 enzyme used to process the erythrocyte includes at least 10 contiguous amino acids of the polypeptide (or the nucleotide sequence encoding same) as specified by: SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, or SEQ ID NO: 8. In certain embodiments, the enzymatic processing of the erythrocyte is performed at from about pH 6.0 to about pH 8.0, preferably about pH 6.5 to about pH 7.5, or more preferably about pH 7.0 to about
15 pH 7.5. In currently preferred embodiments, enzymatic processing of the erythrocyte is performed using 0.01-1000 μ g of enzyme per ml blood cells, preferably 0.1-500 μ g of enzyme per ml blood cells, more preferably 1-100 μ g of enzyme per ml blood cells. Most preferably enzymatic processing of antigens from the erythrocyte preparation is accomplished using 1-10 μ g enzyme/ml blood cells.

20 In yet another aspect, the invention includes a method of modifying a red blood cell, comprising: obtaining a group B or group AB erythrocyte, suspending it in a buffer solution having an approximately neutral pH, and contacting it with an enzyme having at least 10 contiguous amino acids of the polypeptide sequence specified as or encoded by: SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, or SEQ ID NO: 8, thereby
25 cleaving the immunodominant B epitopes on the group B or group AB erythrocyte as determined by serological typing or hemagglutination assays. In one embodiment, the blood cell substantially lacks immunodominant B epitopes as determined by serological typing or hemagglutination assays, and also lacks linear α 1-3 linked Gal structures. In another embodiment, the blood cell substantially lacks immunodominant B epitopes as determined by serological typing or
30 hemagglutination assays, but retains linear α 1-3 linked Gal structures.

 In yet another aspect, the invention includes a method of treating a subject, comprising: identifying a subject in need of type A, O or AB blood, the subject being seropositive for anti-B antibodies; obtaining a modified blood cell preparation of seroconverted B cells, or obtaining the same by the methods described herein, and; transfusing the modified blood cell preparation into
35 the subject, wherein the subject does not immunologically reject the transfused blood cells.

 In yet another aspect, the invention includes a purified polypeptide, the polypeptide having at least 10 contiguous amino acids of the sequences: SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID

NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, or SEQ ID NO: 8, wherein the enzyme displays α 3 galactosidase activity, and a neutral pH optima. In certain embodiments, the purified enzyme, comprises: a polypeptide having at least 10 amino acids of the following sequence numbered accordingly when aligned with SEQ ID. NO:2: M at residue 10; G at residue 47; G at residue 84; Y at residue 86; Y at residue 99; N at residue 102; K at residue 114; T at residue 127; G at residue 130; G at residue 132; G at residue 139; N at residue 156; D at residue 160; P at residue 164; G at residue 205; R at residue 277; R at residue 281; F at residue 287; G at residue 308; Q at residue 312; I at residue 317; R at residue 333; D at residue 340; G at residue 346; G at residue 349; G at residue 360; D at residue 363; D at residue 364; N at residue 367; H at residue 369; G at residue 370; T at residue 371; G at residue 396; E at residue 462; N at residue 463; T at residue 465; T at residue 467; P at residue 468; R at residue 483; G at residue 484; L at residue 486; T at residue 489; N at residue 498; I at residue 508; D at residue 513; W at residue 517; E at residue 519; G at residue 521; D at residue 525; I at residue 528; N at residue 531; F at residue 533; I at residue 549; P at residue 553; I at residue 573; A at residue 590; G at residue 595; N at residue 601; and, I at residue 629; where the polypeptide has at least 20% identity with SEQ ID NO: 2, and where the polypeptide also has α 3 galactosidase activity. In one embodiment, the polypeptide demonstrates specificity for branched alpha galactose structures but not linear alpha galactose structures. In another embodiment, the polypeptide demonstrates specificity for linear alpha galactose structures but not α 1-4 gal structures. In one embodiment, the purified enzyme comprises a polypeptide including nine contiguous amino acids having the sequence DD(P/A)(V/I)N(V/I)HGT (SEQ ID NO: 10). In another embodiment, the purified enzyme comprises a polypeptide including twenty-one contiguous amino acids having the sequence: DXXXW(Y/F)E(S/T)GXXXD(L/V)(L/T)(K/R)XNXF, (SEQ ID NO: 11) where X can be any amino acid. In one embodiment, the purified enzyme includes functional equivalents thereof having α 3 galactosidase activity. In certain embodiments, the polypeptide includes a truncated variant lacking a signal sequence.

In another aspect, the invention includes a method of producing a recombinant enzyme, comprising: obtaining a nucleic acid encoding: SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, or SEQ ID NO: 8; expressing the nucleic acid in a cell transfected therewith; inducing expression of the nucleic acid encoding the enzyme; and purifying the expressed enzyme from the cell. In various embodiments, the invention includes a non-naturally occurring prokaryotic cell, comprising: an expression vector not found in a wild-type prokaryotic cell, the expression vector having a nucleic acid sequence encoding a polypeptide having the sequence specified by: SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, or SEQ ID NO: 8. It will be appreciated that due to the degeneracy of the genetic code, the recombinant enzyme can be optimized for expression in the particular host organism, as is generally known to those skilled in the art.

In a currently preferred embodiment, the invention relates to a family of α -galactosidases, and more particularly to the use of these to catalyze the removal of substantially all of the immunodominant B antigens on B and AB erythrocytes. The most preferred α -galactosidases are those that are active at a neutral pH, and catalyze removal of the immunodominant B antigens on
5 B and AB erythrocytes and not other lipid-linked straight-chain carbohydrate α Gal epitopes such as, P1 antigen (Gal α 1,4 Gal β 1,4 GlcNAc β 1,3 Gal β 1,4 Glc β 1 ceramide) and Pk antigen (Gal α 1,4 Gal β 1,4 Glc β 1 ceramide, also known as globotriosylceramide (Gb3Cer)/CD77). Particular α -galactosidases from this family are described below as, SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8, the fragment SEQ ID NO: 1
10 and the consensus sequence of SEQ ID NO: 9.

In yet another aspect, this invention provides methods for the screening and selection of enzymes with the above described preferred unique characteristics and methods of purification and amino acid sequencing useful for cloning and expression of the genes encoding these enzymes. These methods provide for bacterial isolates producing such preferred enzymes. Such
15 other applications and features of the invention will be apparent from the detailed description that follows.

In yet another aspect, this invention provides compositions and methods for the enzymatic removal of the immunodominant monosaccharides on tissues, such as a tissue for xenotransplantation. Specifically the present invention provides a novel family of α 3-glycosidases
20 (as described above), used for the enzymatic removal of the Galili antigen from non-human animal tissues, thereby converting these to non-immunogenic tissues suitable for transplantation. Examples of an α 3-galactosidase include, but are not limited to, of any of SEQ ID NO: 2-9.

The method of preparing a tissue for xenotransplantation comprises obtaining a tissue from a non-human animal source, incubating the tissue with a polypeptide having
25 α 3-galactosidase activity thereby removing immunodominant α 1-3 linked terminal galactose residues from the tissue, and isolating the tissue from the polypeptide and the enzymatically removed galactose, thereby rendering the tissue suitable for xenotransplantation into humans. In one embodiment, the tissue from a non-human animal source is porcine connective tissue. In another embodiment, the porcine connective tissue is a ligament.

In another embodiment, the tissue from a non-human animal source is an organ including liver, kidney, or heart. In yet another embodiment, the tissue from a non-human animal source is non-immunogenic injectable collagen; bone xenografts; soft tissue and proteoglycan-reduced soft tissue xenografts; xenograft heart valves; meniscal xenografts; and tissue matrices, wherein the tissues are α 1,3-galactose-deficient tissues that have been modified using an α 3-galactosidase.
30 Examples of an α 3-galactosidase include, but are not limited to, of any of SEQ ID NO: 2-9.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 illustrates HPTLC analysis of α -galactosidase activity in the culture supernatant of *Streptomyces griseoplanus* grown in rich media (See Table II for formulations) with the AMC labeled blood group B tetrasaccharide substrate. The fermentation was carried out in for 1 day in YM and 3 days in BP media, at 30 °C, 220 rpm. Assays were performed by mixing equal volumes of the culture supernatant and 0.1 mM B-tetra in 100 mM NaPO₄ (pH 6.8), and incubated at room temperature for 1 hr. One μ L was sampled from each reaction and quickly applied onto HPTLC. Designations: NE, no enzyme control; Ctrl, positive control reaction by using Coffee bean α -galactosidase; CS, culture supernatant of *S. griseoplanus*; S, substrate, *i.e.*, B tetrasaccharide; B-tetra, B tetrasaccharide; H-tri, H trisaccharide; Origin: the position in HPTLC where samples were applied. The TLC plate was developed in chloroform-methanol-water (vol/vol/vol: 60/35/8). The plate was scanned and photographed by a Bio-Rad Fluor-S Multimager with Quantity One - 4.1.1 software.

FIG. 2 illustrates a HPTLC analysis of enzyme assays of *Streptomyces griseoplanus* culture supernatants, recovered from cultures grown in minimal media with 18 different carbon sources, with the AMC labeled blood group B tetrasaccharide substrate. Assays were performed by mixing equal volumes of each culture supernatant and 0.025 mM B-tetra in 100 mM NaPO₄ (pH 6.8), and incubated at room temperature. One μ L was sampled from each reaction at 1 hr and spotted onto TLC plate. The carbon sources, indicated by number 1-18 at the top of the panel, are 18 different sugars used in the fermentation as shown in Table III. Designations: B-tetra: B tetrasaccharide, the substrate; H-tri: H trisaccharide, the product of the B-tetra substrate by α -galactosidase cleavage (The fast moving products above H-tri indicate the presence of fucosidase and β -galactosidase in the culture supernatants that cause further degradation of H trisaccharide into di- and monosaccharides). Fermentations: Cryostocks of *Streptomyces griseoplanus* were thawed and inoculated into YM (~1:5-10, v/v) and incubated at 30°C, 220 rpm, for 24 hrs. The culture was passed onto BP media (~1:20, v/v) and fermentation was continued for 72 hrs. The mycelia, harvested from 100 mL of BP culture by centrifugation, were washed 3 times using basal minimal media (the minimal media lack of carbon source and trace metal/vitamin additives) to eliminate the rich media as much as possible. The pellet was then re-suspended in 100 mL of 2X basal minimal media with additives. The mycelia suspension was then aliquoted into 50 mL conical tubes at 2.5 mL/tube. Different carbon sources and water were then added to a final concentration of 0.5% and a final volume of 5.5 mL. Each carbon source was tested in duplicate. The 36 cultures of 5.5 mL of each, with 18 different carbon sources were incubated at 30 °C, 220 rpm. Aliquot of 0.16 mL of culture was sampled from each tube at 43 and 71 hours.

FIG. 3 illustrates an HPTLC analysis of enzyme assays of culture supernatant of *Streptomyces griseoplanus* grown in minimal media with either galactose or lactose as the sole carbon source with the AMC labeled blood group B tetrasaccharide substrate. Assays were

performed by mixing equal volumes of each culture supernatant and 0.1 mM B-tetra in 100 mM NaPO₄ (pH 6.8), and incubated at room temperature. One μ L was sampled from each reaction at 20 min and applied onto a HPTLC plate for analysis. Designations: B-tetra: B tetrasaccharide, the substrate; H-tri: H trisaccharide, the product of the B-tetra substrate by α -galactosidase cleavage (The fast moving products above H-Tri indicate the presence of fucosidase and β -galactosidase in the culture supernatants that cause further degradation of H-Trisaccharide into di- and monosaccharides); Carbon source: #4, galactose; #7, lactose; NE, no enzyme control; Ctrl, positive control reaction by using Coffee bean α -galactosidase.

FIG. 4 illustrates an HPTLC analysis of the enzyme assay of α -galactosidase activity in the protein solution after passing CEX or DEAE column with B-tetra substrate. About 450 ml of *Streptomyces griseoplanus* supernatant, harvested from 800 mL of culture in an 1 liter fermenter fermentation grown in minimal media with galactose, stored frozen at -80°C , was thawed at for 24 hrs at 4°C and centrifuged for 30 min at 4°C , 20,000 rpm. The recovered supernatant was passed through a 15 mL cation exchange chromatography column (CEX) (Macro-Prep High S Support, BioRad, Cat. #156-0031), pre-equilibrated with 40 mM NaPO₄, 10 mM NaCl (pH 6.8). The flowthrough containing the enzyme activity was collected. The column was washed sequentially with 40 ml of equilibration buffer, 40 mL of the same buffer with a slightly increased pH (7.3). The flowthrough and washes were pooled and loaded directly onto a 2.5 mL DEAE column (DEAE Sepharose, Sigma, Cat. # DEF100) pre-equilibrated with CEX equilibration buffer and the flowthrough was collected. The column was washed with 50 ml of CEX equilibration buffer to remove the residual enzyme from the column. The pooled protein solution of DEAE flowthrough and wash (~600 mL) was concentrated using Centricon Plus 80 Centrifugal filter devices (Millipore Cat.#UFC5LGC02) and buffer-exchanged into 10 mM NaPO₄ (pH 7.0) in the same device to a final volume of 23 mL.

FIG. 5 illustrates HPTLC analysis of α -galactosidase activity in various fractions from Hydroxyapatite step with B-tetra substrate. The protein sample in 10 mM NaPO₄, pH 7.0, was loaded onto a 2.5 mL Hydroxyapatite column (Bio-Gel HT Hydroxyapatite, Bio-Rad Cat. # 130-0150), pre-equilibrated with 10 mM NaPO₄ (pH 7.0). The column was washed with equilibration buffer and washed/eluted stepwisely with increasing amount of NaPO₄ (10 to 100 mM). No activity can be detected in the flowthrough, indicating the effective binding of the enzyme to the column in the presence of 10 mM NaPO₄ (pH 7.0). The appearance of enzyme activity in 30 mM NaPO₄ wash and nearly complete lack of activity in 100 mM NaPO₄ wash indicate simple elution of the enzyme from the Hydroxyapatite column just by using 30-50 mM NaPO₄ (pH 7.0). Designations: Pre, protein solution before being loaded onto the column; FT, flowthrough.

FIG. 6 illustrates HPTLC analysis result of α -galactosidase activity in fractions from Cibacron Blue 3GA step with B-tetra substrate. The pooled activity fractions from Hydroxyapatite step was diluted 1:1 with H₂O and applied onto a 2.5 mL of Cibacron Blue column (Cibacron Blue 3GA, Sigma, Cat. # C-1285), equilibrated with 10 mM Tris (pH 7.5). The column was washed with

the equilibration buffer and further washed/eluted with equilibration buffer with increased amount of salt as indicated at the bottom of the panel. The enzyme activity was distributed between 100 and 400 mM NaCl washes. Designations: Pre, protein solution before being loaded onto the column; FT, flowthrough.

5 FIG. 7 illustrates HPTLC analysis of α -galactosidase activity in various fractions from AEX step with B-tetra substrate. The pool of enzyme activity fractions from Cibacron Blue was concentrated and buffer-exchanged into 40 mM Tris, 10 mM NaCl (pH 8.5), to a final volume of 3.7 mL. The protein solution was loaded onto an 1 mL of AEX column (Macro-Prep High Q Support, Bio-Rad Cat. # 156-0051), pre-equilibrated with 40 mM Tris, 10 mM NaCl, pH 8.5. The column was first washed with equilibration buffer and then washed/eluted with the same buffer containing increasing amount of salt as indicated at the bottom of the panel. Designations: Pre, protein solution before being loaded onto the column; FT, flowthrough; Washes/Elutes, column wash and/or elution samples; [NaCl] (mM), the salt concentration in the wash/elution buffer; Fraction #, fractions collected in each wash/elution step; B-tetra, B tetrasaccharide, the substrate; 10 H-tri, H trisaccharide, the product (the faster moving product above H-tri indicates the presence of contaminating fucosidase activity in the protein sample that cause further degradation of H trisaccharide into disaccharride). 15

 FIG. 8 illustrates a SDS-NuPAGE (Novex 4-12% Bis-Tris Gel with MOPS buffer, stained with SilverQuest Silver Staining kit, Mark12 Unstained Standard, all Invitrogen products) analysis of *Streptomyces griseoplanus* α -galactosidase activity purified by AEX. The HPTLC analysis of enzyme assays of fraction #3 of the wash/elution sample at each salt concentration in the AEX step as shown in FIG. 7, was placed at the top of the gel for easy comparison of the enzyme activity and protein band (s) on the gel. A single protein band, ~ 70 kDa, indicated by an arrow at right side of the panel labeled with putative α -galactosidase, is shown in the peak α -galactosidase activity. Designations: B-tetra, B tetrasaccharide, the substrate; H-tetra, H trisaccharide, the product. 20

 FIG. 9 illustrates comparative analyses of S12 chromatography fractions of partially purified *Streptomyces griseoplanus* α -galactosidase by SDS-NuPAGE and enzyme activity assay using B tetrasaccharide, analyzed by HPTLC. The putative 70 kD α -galactosidase band is indicated by an arrow at right side of the panel using Rainbow Molecular Weight Marker (Amersham, Cat. # RPN800). Designations: Ctrl, NEB A-zyme of known concentration; B-tetra, B tetrasaccharide, the substrate; H-tri, H trisaccharide, the product. 30

 FIG. 10 illustrates the alignment of the peptide obtained by Edman sequencing of HPLC fractionated trypsin digest of a novel *Streptomyces griseoplanus* α -galactosidase (SEQ ID NO: 1), with a hypothetical protein from *Streptomyces avermitilis* (GenBank access # BAC74979.1, GI:29610934, SEQ ID NO: 2). The amino acids in SEQ ID NO: 2 that correspond to those of SEQ ID NO: 1 are underlined. The alignment was obtained by blast analysis of the peptide using 35

"search for short, nearly exact matches" against NCBI nr database [(Score = 51.5 bits (114), Expect = 3e-06; Identities = 18/29 (62%), Positives = 24/29 (82%), Gaps = 0/29 (0%)]. The amino acid sequence is shown in a single-letter code. The identical residues are indicated by bold capital letters, similar residues by plain bold letters, and different residues by small letters.

5 FIG. 11 illustrates an HPTLC analysis of enzyme assays of culture supernatant and pellet lysate of *Streptomyces avermitilis* grown in YM media (See Table II for formulations) with the AMC labeled blood group B tetrasaccharide and 4-methylumbelliferyl α -D-galactopyranoside (α -Gal pNP) substrates. The fermentation was carried out in for 3 days at 30°C, 220 rpm. Assays were performed by mixing equal volumes of the culture supernatant and 0.1 mM B-tetra or 0.5 mM of

10 α -Gal pNP in 100 mM NaPO₄ (pH 6.8) and incubated at room temperature overnight. One μ L was sampled from each reaction and quickly applied onto HPTLC. Designations: NE, no enzyme control; Ctrl, positive control reaction by using Coffee bean α -galactosidase; CS, culture supernatant of *S. avermitilis*; PT, pellet lysate; B-tetra, B tetrasaccharide; H-tri, H trisaccharide; MU, 4-methylumbelliferone, the cleavage product of α -Gal pNP; Origin: the position in HPTLC

15 where samples were applied. The TLC plate was developed in chloroform-methanol-water (vol/vol/vol: 60/35/8). The plate was scanned and photographed by a Bio-Rad Fluor-S Multimager with Quantity One -4.1.1 software.

 FIG. 12 illustrates protein sequence alignment of the putative novel α -galactosidase from *Streptomyces avermitilis* (SEQ ID NO: 2) with a number of the first protein hits of unknown

20 functions by blasting SEQ ID NO: 2 against NCBI nr databases. The alignment was performed using CLUSTALW multiple alignment (NPS@: Network Protein Sequence Analysis, TIBS 2000: 25;147-150, Combet C., Blanchet C., Geourjon C. and Deléage G.). Alignment data: Alignment length : 665; Identity (*) : 59 is 8.87 %; Strongly similar (:): 86 is 12.93 %; Weakly similar (.) : 42 is 6.32 %; Different : 478 is 71.88 %. The sequences are as follows: SA (625 residues SEQ ID

25 NO: 2); BT α (568 residues SEQ ID NO: 3); BF α 1 (605 residues SEQ ID NO: 4); BF α 2 (605 residues SEQ ID NO: 5); BF β 1 (595 residues SEQ ID NO: 6) ; BF β 2 (595 residues SEQ ID NO: 7); BT β (615 residues SEQ ID NO: 8). SEQ ID NO: 9 is a consensus sequence of the sequences SEQ ID NOs: 2-8. Designations: SA, BT and BF, the putative α -galactosidases from *Streptomyces avermitilis* MA-4680, Bacteroides thetaotaomicron VPI-5482 and Bacteroides

30 fragilis, respectively; α and β : 2 different copies of α -galactosidases from B. thetaotaomicron VPI-5482; α 1 and β 1: 2 different copies of α -galactosidases from B. fragilis YCH46; α 2 and β 2: two different copies of α -galactosidases from B. fragilis NCTC 9343.

 FIG. 13 illustrates an HPTLC analysis of the enzyme activities, of whole cell lysates of the cell pellet from IPTG induced cultures of *E. coli* clones (containing plasmids expressing the recombinant α -galactosidase gene from *Streptomyces avermitilis*), with the AMC labeled blood group B tetrasaccharide substrate. One mL of TB medium with antibiotics (48.2 g of EZmix Terrific Broth, Sigma T-91790, 8 mL of glycerol, 34 mg of Chloamphenicol, and 30 mg of Kanamycin per

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liter medium) was added to each 1.5 mL microtube containing the agar plug carrying a single colony. The cap was closed and incubation was performed overnight at 37°C, 250 rpm. One half mL of an overnight culture was inoculated into 10 mL of medium in a 50 mL conical tube and the incubation was carried under the same conditions. The cell density reached 0.3-0.6 OD_{600nm} in about 2 hrs at 220 rpm. The culture was removed from the shaker and kept at room temperature for ~20 min. Meanwhile, the temperature of the incubator was lowered to ~26°C. IPTG was then added to each culture at a concentration of 0.1 mM and all cultures were re-placed in the shaker and agitated at 220 rpm, to start protein induction. A 0.5 mL aliquot was removed aseptically from each tube in 1 hr and the cells pelleted with a bench top centrifuge at the highest setting for 5 min.

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Twenty μ L of lysis buffer (0.9 mL of 40 mM NaPO₄, 10 mM NaCl, pH 6.8, 0.1 mL of BugBuster 10X, Novagen 70921-4, 1 mg lysozyme/mL, and 5 μ L of benzonase, Novagen 70664-3, per milliliter lysis buffer) was added to each tube to suspend the pellet and lyse the cells, which was assisted by pipetting the suspension up and down a few times. The lysis was completed in 5-10 min. An aliquot (2.2 μ L) of the crude whole lysate was analyzed subsequently by mixing with equal volume of a substrate solution containing 0.1 mM of B-tetra in 100 mM NaPO₄ (pH 6.8) and incubated at room temperature. One μ L of the digestion was removed in 10 min and spotted onto a HPTLC plate. Designations: Ctrl, positive control reaction by using Coffee bean α -galactosidase; 1 and 2: whole lysates from two individual colonies of the same construct expressing the full length novel galactosidase from *Streptomyces avermitilis*; B-tetra, B tetrasaccharide; H-tri, H trisaccharide; Origin: the position in HPTLC where samples were applied. The TLC plate was developed in chloroform-methanol-water (vol/vol/vol: 60/35/8). The plate was scanned and photographed by a Bio-Rad Fluor-S Multimager with Quantity One -4.1.1 software. FIG. 14 confirmed that the novel B-zyme can be efficiently expressed in *E. coli*, but as inclusion bodies. Therefore, expression needs to be optimized or an efficient refolding method needs to be developed for the application of this novel B-zyme.

FIG. 14 illustrates a SDS-NuPAGE analysis of α -galactosidase (SEQ ID NO: 2) expressed in *E. coli* (Novex 4-12% Bis-Tris Gel with MOPS buffer, stained with Colloidal Blue Staining kit, Mark12 Unstained Standard, all Invitrogen products). The lysate from each culture was prepared similarly as described in FIG. 13 legend with an increased scale. An aliquot of the each whole lysate was centrifuged at 14,000 g for 5 min. at RT. The supernatant was removed. Twelve μ L of whole lysate or supernatant was mixed with 4 μ L of 4X LDS buffer, supplemented 10% (v/v) β -mercaptoethanol. The pellet was suspended in 1 X LDS sample buffer, supplemented with 2.5% (v/v) β -mercaptoethanol, at a ratio of 16 μ L sample buffer / 12 μ L of whole lysate. All samples were heated at 70°C for 10 min for SDS-NuPAGE analysis. Designations: WL, whole lysate; Sup, supernatant; PT, pellet; U, sample prepared from un-induced culture; I, sample prepared from induced culture.

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FIG. 15 illustrates Multiple ClustalW (BoxShade 3.21) protein sequence alignment of the putative novel α -galactosidase family to identify conserved regions for the design of degenerate

primers. Identical residues and conserved substitutions are highlighted in black and dark gray. The aligned sequences are from *S. avermitilis* MA-4680, *B. thetaiotaomicron* VPI-5482 and *B. fragilis* NCTC 9343. The two sequences from *B. fragilis* YCH624, nearly identical to those from *B. fragilis* NCTC 9343, are not included. The conserved regions used to design a pair of
5 degenerate primers to clone the partial α -galactosidase gene from *S. griseoplanus* 2357 are indicated with a forward arrow for forward primer and backward arrow for reverse primer.

FIG. 16 illustrates protein sequence of a *S. griseoplanus* α -galactosidase highlighting the regions corresponding to the primers used for cloning. The forward and reverse primers are colored dark and light gray respectively. Degenerate primers are underlined.

10 FIG. 17 illustrates a HPTLC analysis of enzyme assays of purified recombinant FragB α -galactosidase with a panel of oligosaccharides of diverse structures. Reactions were performed at room temperature with 10 nmole of substrate and 21 ng of enzyme in 10 μ L of 10 mM NaPO₄, pH 6.8/2.5 mM NaCl, supplemented with 0.25 mg/mL of BSA. One μ L of enzyme assays was removed at desired time points and spotted onto a silica gel-coated TLC plate (EMD Chemicals,
15 NJ), which was developed in chloroform-methanol-water (vol/vol/vol: 30/60/10) for 15 min and product developments were detected by Orcinol/H₂SO₄ staining. Designations: Reaction time for each substrate from left to right: 0 (sampled from control reaction containing no enzyme), 5, 10, 20, 40 and 80 min. Detailed structures of the substrates are described in Table V. The cleavage of the substrate resulted increased migration as observed for B-tri, B-di and linear B, but not for P₁,
20 P^k and A-tri, indicating the lack of cleavages.

FIG. 18 illustrates a HPTLC analysis of enzyme assays of purified recombinant FragB B-zyme with AMC-B-tetra under different pH. Reactions were performed at room temperature with 1 nmole of substrate and ~8 ng of enzyme in 10 μ L of buffer at pH 2.0 to 9.0, supplemented with 0.25 mg/mL of BSA. One μ L of enzyme assays was removed at desired time points and spotted
25 onto a HPTLC plate, which was developed in chloroform-methanol-water (vol/vol/vol: 60/35/8). The plate was scanned and photographed by a Bio-Rad Fluor-S Multilmager with Quantity One – 4.1.1 software. The 1 X buffers used in the reactions were derived from 2 X buffers described as follows: pH 2.0, 0.1 M citric acid; pH 2.5 – 5.5, 0.1 M citric acid/0.2 M Na₂HPO₄; pH 6.0-7.5, 0.2 M NaH₂PO₄/0.2 M Na₂HPO₄; pH 8.0-9.0, 0.2 M Tris/HCl. Assay mixtures were sampled at 5 (top panel) and 10 min (bottom panel). Designations: B-tetra, B tetrasaccharide; H-tri, H trisaccharide; Origin: the position in HPTLC where samples were applied.
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FIG.19. illustrates the analysis of the enzyme activity under different pH using chromogenic para-nitrophenyl derivative, Gal α -pNP. Assays were carried out by using 2.5 mM substrate, 8.5 μ g enzyme in 400 μ L of buffers between pH 2.0 – 9.0 as described in the legend of
35 FIG.17, at 26°C for 5 min, terminated with 600 μ L of 1.0 M Na₂CO₃ and, read at 405 nm. A molar extinction coefficient of 18,300 was used to calculate the amount of released nitrophenol. One unit was defined as the amount of enzyme required to cleave 1 μ mole of substrate per minute under

the experimental condition. The specific activity at each pH was then calculated and plotted versus pH.

FIG. 20 illustrates the substrate specificity of *Bacteroides fragilis* α -galactosidases. Enzyme assays were carried out without enzyme (–) and with ~30 ng enzyme (+), ~1.0 mM substrate in 10 μ L of 10 mM NaPO₄, pH 6.8, 2.5 mM NaCl, supplemented with 0.25 mg/mL BSA. Reactions were monitored by TLC during incubation at 26°C and the 2 hr time point is shown. Cleavages of the branched blood group B trisaccharide (B-tri) to the H disaccharide (H-di) by BF α 2 (FragA) α -galactosidase, and all B structures by BF β 1 (FragB) α -galactosidase, were complete within 5-20 min (not shown), whereas no cleavage of other oligosaccharide substrates were detected after 2 hr incubation. The TLC plates were developed in chloroform/methanol/water (30/60/10, v/v/v) for 15 min and stained by heating with 0.05% Orcinol in 0.5M H₂SO₄.

DETAILED DESCRIPTION OF THE INVENTION

This invention is directed to the development and application of a screening and selection strategy for novel α -galactosidases with preferred specificities for the blood group B structures, and with preferred performance in the enzymatic conversion of blood products and animal tissues, over an approximately neutral pH range. Table 1 lists the complex structures of antigens found on blood cells.

For the purpose of this invention, blood group B active oligosaccharide derivatives were synthesized or produced by enzymatic removal of α Gal from various substrates. Furthermore, glycosphingolipids with structures 3, 6, 21, and 25 were purified from human erythrocytes or produced therefrom by glycosidase treatments as previously described (Clausen *et al.*, Proc. Natl. Acad. Sci. USA 82(4): 1199-203, 1985, Clausen *et al.*, J Biol Chem. 261(3): 1380-7, 1986, Clausen *et al.*, Biochemistry 25(22): 7075-85, 1986, Clausen *et al.*, J Biol Chem. 262(29): 14228-34, 1987). Thin-layer chromatography assays to quantitatively determine removal of α Gal or α GalNAc from the AMC derivatives or glycosphingolipids were developed.

Preferred α -galactosidases have high substrate specificity for blood group B branched saccharide structures, a generally neutral pH optima and can be produced cost-effectively as recombinant proteins in unicellular organisms such as bacteria and yeast. Our prior patent application (U.S.S.N. 10/251,271), developed a screening assay for the preferred enzyme activities using B tetrasaccharide AMC derivative substrates, and measured enzyme activities at neutral pH. Further, activities were compared to activities using p-nitrophenyl monosaccharide derivatives in order to identify activities with preference or exclusivity for the complex substrates. In that application, we disclose the use of this screening assay on a large panel of bacterial and fungal isolates (3100), and therein we identified several bacterial isolates expressing α -N-acetyl galactosaminidase or α -galactosidase activities measured with A or B tetrasaccharide AMC

substrates, but no or insignificant levels of activity with the corresponding p-nitrophenyl monosaccharide substrates. One of each of these activities was further analyzed after sero- and genotyping these as *Streptomyces* strains. Analysis of strain #8 was determined to have α -N-acetylgalactosaminidase activity revealed that the activity was insoluble and was associated with the cell mass. Strain #8 was deposited on February 14, 2002 with the American Type Culture Collection (ATCC) and has been assigned ATCC Deposit No. PTA-4076. In contrast, strain #2357 was determined to have α -galactosidase activity, and the activity was determined to be soluble, found in the supernatant of transformed cells lysed by French press. Strain #2357 was deposited on February 14, 2002 with the American Type Culture Collection and was assigned ATCC Deposit No. PTA-4077. Because it is considerable simpler to purify a soluble protein, we chose to initially purify and sequence the protein from strain #2357.

The enzyme that we found in the soluble fraction of strain #2357 was partially purified. Detailed analysis of the substrate specificity of the partially purified α -galactosidase demonstrated an unprecedented fine specificity for the branched B blood group structures, but no linear structures capped by α 1-3 or α 1-4 galactose residues were cleaved by this enzyme. Analysis of its pH optimum showed the preferred conditions to be pH 5.5 to 7.0. The identified α -galactosidase activity is therefore highly preferred over enzymes known in the prior art due to its restricted substrate specificity, high specific activity for group B structures, and pH optimum. SDS-PAGE analysis of the resulting partially purified crude extract revealed 3-4 protein bands in the 40-80 kDa region having the α -galactosidase activity. Gel filtration analysis of the preparation showed the activity migrated comparable to BSA, indicating a globular protein having a molecular weight of about 40-80 kDa. A single short sequence was obtained:
Phe-Ala-Asn-Gly-Leu-Leu-Leu-Thr (SEQ ID NO: 1).

Subsequent to these studies, and as disclosed in the present invention, we have discovered a new family of polypeptides, having α -galactosidase activities, and have developed methods for their induction, purification, sequencing and cloning. As discussed below, the polypeptide family is distinct from the previously partially purified protein from strain #2357, and notably these family members do not contain the sequence shown as SEQ ID NO: 1. The new induction strategy involves growth of the appropriate bacterium on defined carbon sources and minimal medium, which results in a significant increase in production of the α -galactosidase polypeptides. Known α -galactosides (generally having an acidic pH optimum and substrate specificity for Gal α -pNP or other simple monosaccharides) are not secreted in *griseoplanus* and related *Streptomyces* strains under the same growth conditions that produces these novel polypeptides.

The present invention therefore provides a novel method for the recombinant expression and purification of certain α -galactoside polypeptides. This purification strategy applied in combination with the novel growth and induction methods resulted in successful purification to

apparent homogeneity of the α -galactosidase polypeptides in sufficient quantities for amino acid sequencing, and blood product and tissue conversions.

The following successive steps were used to achieve purification to apparent homogeneity: cell broth supernatant derived from cultures of *S. griseoplanus* #2357 was first
5 passed unbound successively through CEX and DEAE columns (FIG. 4). Subsequently the activity was bound and eluted successively on a Hydroxyapatite column (FIG. 5), a Cibacron Blue column (FIG. 6), and finally an AEX column (FIGs. 7 and 8). Throughout the purification scheme the protein was followed by an analysis of its enzymatic activity in various fractions; the final protein product was also analyzed by SDS-NuPAGE. Identification of the α -galactosidase protein was
10 obtained by comparison of protein banding pattern by SDS-NuPAGE silver staining and AEX and S12 gel filtration chromatographies (FIGs. 8-9). Only one band migrating as 70 kD by SDS-NuPAGE and S12 gel filtration, corresponded with the observed α -galactosidase activity. The protein identified as described, was finally separated by NuPAGE gel electrophoresis and the Coomassie stained 70 kD band was cut out of the gel and submitted for amino acid sequence
15 analysis. Internal amino acid sequence information was obtained by mass spectrometric analysis (MALDI-TOF) and Edman degradation after trypsin digestion. None of the short sequences obtained showed high degree of identity with known sequences in public databases (GenBank). A Blast database search of a 30 amino acid peptide (the longest peptide sequence obtained by internal sequencing and confirmed by MS/MS, using "Search for short, nearly exact matches")
20 identified a putative open reading frame predicted to encode a protein (SEQ ID NO: 2) from the genome sequence of *Streptomyces avermitilis* (GenBank access # BAC74979.1, GI:29610934). The complete genome of *Streptomyces griseoplanus* is not available and no related sequences derived from this genus were identified in database searches. *Streptomyces avermitilis* and *Streptomyces griseoplanus* are closely related. We therefore tested if *Streptomyces avermitilis*
25 also contained the identified α -galactosidase activity, which was previously demonstrated to be very rare among bacterial isolates including many *Streptomyces* isolates.

Streptomyces avermitilis (ATCC 31267) culture supernatant was assayed for secreted α -galactosidase and as shown in FIG. 11, clear evidence of the presence of α -galactosidase activities in both culture supernatant and pellet lysate was observed, as determined by digestion of
30 B-tetra oligosaccharide substrates. However, the cleavage of a simple substrate (α -Gal pNP) by the secreted *Streptomyces avermitilis* α -galactosidase was negligible. In contrast, complete cleavage of α -Gal pNP was observed for the *Streptomyces avermitilis* α -galactosidase obtained from the cellular fraction. Therefore, the secreted and cellular α -galactosidases are probably not of the same identities. Part of the secreted galactosidase is likely to be the novel α -galactosidase
35 that prefers branched substrate to simple (linear) substrates, while most of the cellular α -galactosidase activities if not all are observed to have conventional glycosidase activities. The similarities of the polypeptide from *S. avermitilis* (SEQ ID NO: 2) to the α -galactosidase from *S. griseoplanus* with respect to secretion into culture broth, predicted molecular weight, and the

sequence similarities with *S. griseoplanus*, indicates that the *S. avermitilis* protein represents a homologue of the originally identified *S. griseoplanus* derived α -galactosidase.

The identified from *S. avermitilis* polypeptide (SEQ ID NO: 2) consists of 625 amino acids and showed no significant similarity to any other known proteins. Back searches with SEQ ID NO: 2 identified several novel protein sequences (SEQ ID NO: 3-8) from exclusively prokaryotic genomes, with sequence similarities shown as follows in Table 1A:

Table 1A
Identity (overall %) of *Bacteroides* α -Galactosidases to the *S. avermitilis* Enzyme.

SEQ ID NO.	GI NO.	Abbr.	Amino Acids	Identity to SA (%)
2	gi 29833810 ref NP_828444.1 <i>S. avermitilis</i>	SA	625	100
3	gi 29340474 gb AAO78266.1 <i>Bacteroides thetaiotaomicron</i> VPI-5482	BT α	568	30.35
4	gi 53715733 ref YP_101725.1 <i>Bacteroides fragilis</i> YCH46	BF α 1	605	30.23
5	gi 60495103 emb CAH09922.1 <i>Bacteroides fragilis</i> NCTC 9343	BF α 2	605	29.30
6	gi 60491830 emb CAH06588.1 <i>Bacteroides fragilis</i> NCTC 9343	BFB1	595	24.30
7	gi 53712216 ref YP_098208.1 <i>Bacteroides fragilis</i> YCH46	BFB2	595	24.30
8	gi 29341569 gb AAO79356.1 <i>Bacteroides thetaiotaomicron</i> VPI-5482	BT β	615	22.82

All polypeptides identified were analyzed by multiple sequence alignments as shown in FIG. 12; a consensus sequence is provided as SEQ ID NO: 9. These polypeptides represent a new family of novel α -galactosidases, that have unique substrate specificity described in more detail below, and having the common feature of an approximately neutral pH optima.

The gene sequences for these members of this α -galactosidase family have allowed development of recombinant expression systems for these polypeptides, using a variety of prokaryotic or eukaryotic cells and expression systems, and permit purification of recombinant forms of these enzymes using established protein purification procedures (for example HIS tag expression and purification systems).

EXAMPLES

Enzyme assays:

Substrates consisting of a series of complex blood group ABH oligosaccharide structures, such as 7-amino-4-methyl-coumarin derivatives were custom synthesized by Alberta Chemical Research Council (see, U.S.S.N. 10/251,271). Other substrates were available from different suppliers (Sigma-Aldrich). All reagents used were of analytical grade or higher. Standard enzyme assays were performed as follows with the different substrates.

Typical assays were performed by the following procedure: Protein samples were incubated with AMC labeled oligosaccharide at 0.05 mM concentration, with MU-labeled monosaccharide at 0.25 mM concentration, in 2.2-10 μ L reaction in 50 mM NaPO₄ (pH 6.8) for the desired time at 26°C or room temperature. One μ L aliquot was taken at various time points and spotted onto HPTLC to follow product development. The TLC plate was developed in chloroform-methanol-water (vol/vol/vol: 60/35/8). The plate was scanned and photographed by a Bio-Rad Fluor-S Multilimager with Quantity One -4.1.1 software. One unit of enzyme activity is defined as the amount of enzyme required to cleave 1 μ mole of substrate per minute under the experimental conditions.

10 *Fermentations:*

The formulations of various media were listed in Table II. Fermentations in flask and 50 mL conical tubes were performed under standard conditions: 30°C, 220 rpm for the desired length of time. The fermentation was performed at pH 6.8, 30°C, 300-600 rpm, DO = 50%.

15 **Table II.**
Media¹ formulations for growing *Streptomyces griseoplanus* for the production of α -galactosidase polypeptides

YM Medium	
Components	g/L
Yeast extract	3
Malt extract	3
Bacto Soytone	5
Glucose	10
BP Medium	
Components	g/L
Bacto Soytone	15
Malt extract	5
Yeast extract	5
Pharmamedia	5
KH ₂ PO ₄	1
MgSO ₄ ·7H ₂ O	1
CaCO ₃	2.5
Glucose	25
N-Acetylglucosamine ²	0.1

Minimal Medium

Components	g/L
(NH ₄) ₂ SO ₄	2
MgSO ₄ ·7H ₂ O	0.6
0.2M NaH ₂ PO ₄ / K ₂ HPO ₄ pH 6.8	7.5 mL
CaCl ₂	0.1
ZnSO ₄ · 7H ₂ O	0.1
FeSO ₄ · 7H ₂ O	0.1
MnCl ₂ · 4H ₂ O	0.1
Carbon source ²	5
Trace mineral supplement (ATCC Cat. # MD-TMS) ²	10 mL
Vitamins supplement (ATCC Cat. # MD-TMS) ²	10 mL

1. All media without the indicated components were sterilized at 121°C for 25 min.
2. Components were sterilized by 0.22 µm filtration and added to the desired recipe after sterilization.

EXAMPLE 1: INDUCTION OF α -GALACTOSIDASE EXPRESSION IN *STREPTOMYCES GRISEOPLANUS*

5 *S. griseoplanus* was shown in the past to be capable of producing a secreted novel α -galactosidase when grown in proper media, although this enzyme was never purified to homogeneity. A cryostock of this microorganism was inoculated into 5 to 10 volumes of YM media and grown for 24 hrs at 30°C, 220 rpm, in shaking flasks or 50 mL conical tubes depending on the scale of the culture. The YM culture was then inoculated into about 20 volumes of BP

10 media for continuing incubation under the same conditions, to induce production of the galactosidase. The enzyme activity associated with the culture usually peaks in 3 days. The culture supernatant containing the enzyme activity was harvested by centrifugation. FIG. 1 shows a HPTLC analysis of the enzyme assay of a typical spent culture media supernatant with the substrate B-tetra. The identified α -galactosidase was expressed in a very low volumetric yield

15 both in total lysates of cells as well secreted into the medium (approximately ~0.1 U/L culture as analyzed by the B-tetra AMC enzyme assay described under general methods). Therefore, it was largely impossible to isolate sufficient amount of pure protein for sequencing (see, U.S.S.N. 10/251,271). The low expression level of the desired protein, the heterogeneity and protein richness of the rich media, were considered to represent the main factors for the difficulty to purify

20 enough activity for protein identification.

It was considered necessary in the present study, to develop a strategy to induce expression and secretion of the enzyme to achieve a higher starting specific activity. One approach was to use an alternative carbon source instead of glucose. Another approach was to reduce the complexity of the media by using homogeneous media with little organic materials in

25 particular the protein content, *i.e.*, the minimal media. The isolation of the enzyme activity from

such media was expected to be easier and yields of enzyme at each step were expected to be increased.

Considering that the growth of the microorganism in minimal media is very slow and the sensitivity of α -galactosidase production to the growth media composition, *S. griseoplanus* was first grown in rich media following the standard protocol, *i.e.*, 24 hrs in YM, 72 hrs in BP. The mycelia were then harvested from the culture by centrifugation. The pelleted mycelia were washed thoroughly with basal minimal media (minimal media lack of carbon source and additives) to eliminate the residual rich media as much as possible. The mycelia pellet was then re-suspended in minimal media lacking a carbon source, which can be easily distributed for carbon source screening as detailed in FIG. 2. The small scale tube cultures were performed under standard fermentation condition. Small aliquots were sampled at different time points and supernatants were recovered for α -galactosidase analysis. A total of 18 carbon sources were studied as shown in Table III. The HPTLC analysis of enzyme assays of culture supernatants with B-tetra are shown in FIG. 2. The complete disappearance of the substrates using 70 hrs fermentation supernatants in lane 4 and 7 clearly distinguish galactose and lactose from other carbon sources in their ability to produce the α -galactosidase. Under current assay condition, 25 pmol of substrate/ μ L of protein sample, 50 mM NaPO₄ (pH 6.8), 1 hr at room temperature, the volumetric yield of the α -galactosidase activity can be calculated as follows:

$$25 \text{ pmol}/(1 \mu\text{L} * 60 \text{ min}) \approx 0.4 \text{ mU/mL or } 0.4 \text{ U/L.}$$

The yield is much higher than a typical yield obtained from rich media culture (~0.1 U/L). Furthermore, the yield is probably underestimated since lack of time point before 1 hr may have missed the end point of the reactions. The preliminary result shows great potential of using minimal media to facilitate α -galactosidase production and purification.

To confirm the remarkable observation using minimal media, we re-evaluated the leading carbon source galactose and lactose for the novel α -galactosidase induction. FIG. 3 shows the HPTLC analysis of reaction assays of fermentation samples taken at different time points grown in minimal media with galactose and lactose as carbon sources. About 90% of the substrate was cleaved by 3 day culture samples in 20 min, which translated to about 4 U/L of culture supernatant. Therefore, as shown in FIG. 3, galactose (lane #4) and surprisingly lactose (lane #7) induced significant α -galactosidase activity. Then conditions for growth and induction using galactose identified above were used without further optimization to develop large-scale fermentations of the *Streptomyces* strain #2357 for isolation of the enzyme. As will be evident from the following examples these conditions were essential for the successful isolation and identification of the resultant α -galactosidase protein, which was different from the enzyme originally disclosed by U.S.S.N. 10/251,271.

Table III. Carbon sources used for screening novel α -galactosidase induction from *Streptomyces griseoplanus* using minimal media.

Carbon Source #	Carbon Source
1	Carob tree crenel flour
2	Dextrin from potato starch
3	D(-) Fructose
4	D(+) Galactose
5	D(+) Glucosamine
6	Glycerol
7	D(+) Lactose monohydrate
8	Malt extract
9	D(+) Maltose monohydrate
10	D-Mannitol
11	D(+) Mannose
12	D(+) Raffinose
13	L(-) Sorbose
14	Starch
15	Sucrose
16	Xylitol
17	D(+) Xylose
18	D(+) Glucose

EXAMPLE 2: PURIFICATION OF A α -GALACTOSIDASE EXPRESSED IN *STREPTOMYCES GRISEOPLANUS* STRAIN #2357

5 A new purification strategy was developed for the novel enzyme since the starting material was substantially different than that used for the partial purification described previously (see, U.S.S.N. 10/251,271). The following steps were used to achieve purification to apparent homogeneity: cell broth supernatant (450 ml), derived from 800 mL of culture carried out in an 1L fermenter as described in Example 1, was subjected to 30 min's high speed centrifugation at

10 20,000 rpm, 4°C. The supernatant was applied to a 15 ml CEX column (Macro-Prep High S support, BioRad, Cat. #156-0031), pre-equilibrated with 40 mM NaPO₄, 10 mM NaCl (pH 6.8), and washed with 40 ml of the equilibration buffer and 40 mM PO₄, 10 mM NaCl (pH 7.3), respectively. The flowthrough and the two washes containing the α -galactosidase activity were pooled (FIG. 4, panel A), and applied onto a second column of 2.5 mL DEAE (DEAE Sepharose,

15 Sigma, Cat. # DEF100) pre-equilibrated with 40 mM NaPO₄, 10 mM NaCl (pH 6.8). The column was then washed with 50 ml of the equilibration buffer. A total of 600 ml containing the α -galactosidase activity was collected from the flowthrough and the wash (FIG. 4, panel B). They were pooled, concentrated with a Centricon Plus 80 Centrifugal filter devices (Millipore Cat. # UFC5LGC02), and buffer-exchanged to 10 mM NaPO₄ (pH 7.0) in the same device to a final

20 volume of 23 mL.

The 23 ml buffer-exchanged sample was applied to a 2.5 ml Hydroxyapatite column (BioRad, Cat. #103-0150) pre-equilibrated with 10 mM NaPO₄ (pH 7.0). The column was washed with 5 ml of the equilibration buffer, and the α -galactosidase activity was eluted stepwise with a NaPO₄ gradient buffer (10 mM/step) from 20 to 100 mM (pH 7.0). The α -galactosidase activity
5 eluted in fractions with 30-50 mM NaPO₄ (FIG. 5). The active fractions were pooled and diluted 1:1 with H₂O and applied to a 2.5 ml Cibacron Blue column (Sigma, Cat. # C-1285) pre-equilibrated with 10 mM Tris (pH 7.5). The column was washed with 10 ml of 10 mM Tris (pH 7.5) and 5 ml of 10 mM Tris, 80 mM NaCl (pH 7.5). The α -galactosidase activity was eluted with 25 ml of elution buffer containing 10 mM Tris (pH 7.5) with increased amount of salt (FIG. 6). The
10 enzyme eluate was concentrated and buffer-exchanged into 40 mM Tris, 10 mM NaCl (pH 8.5) with Centricon YM10 centrifugal filter devices (Millipore Cat. #4205) to a final volume of 3.7 mL. Finally, the buffer-exchanged eluate was applied to a 1 ml AEX column (BioRad, Cat. #156-0031), pre-equilibrated with 40 mM Tris, 10 mM NaCl (pH 8.5). The column was washed with 5 ml equilibration buffer and the α -galactosidase activity was eluted with a NaCl gradient in 40 mM Tris
15 (FIG. 7).

Analyses of the eluate fractions of the AEX column were performed by SDS-NuPAGE and a single band with apparent molecular weight of 70 kD was observed after silver staining (SilverQuest, Invitrogen, Cat. #LC6070) (FIG. 8). Further verification of the identity of the isolated α -galactosidase was provided by gel filtration chromatography. A S12 column (Superose 12™,
20 Amersham, Cat.#17-5173-01) was equilibrated and run with 150 mM ammonium acetate. Partially purified α -galactosidase as described above was applied (250 μ l volume) and 45 fractions (0.5 ml/fraction at 1 ml/min flow rate) were collected (FIG. 9). Fractions #19-21 contained the major protein peak (uv 280 nm). Fractions 19-22 were analysed for α -galactosidase with B-tetra AMC and 10 μ l of each was analyzed by a 4-12% gradient SDS-NuPAGE using 10 and 20 ng of NEB A-
25 zyme as controls (lane 2 & 3). As shown in FIG. 9 the peak α -galactosidase activity correlates fully with the 70 kD band by SDS-PAGE.

EXAMPLE 3: AMINO ACID SEQUENCING OF PURIFIED α -GALACTOSIDASE FROM *STREPTOMYCES GRISEOPLANUS* STRAIN #2357

Approximately 1 μ g α -galactosidase protein as estimated by NuPAGE was prepared as
30 described in Example 2. The protein was separated by 4-12% NuPAGE and stained with Colloidal Blue Staining Kit (Invitrogen, Cat.# LC6025). After destaining of the gel with H₂O, the stained 70 kD bands were excised and washed with HPLC grade H₂O and 50% acetonitrile in H₂O. The sliced gel was subjected to direct sequence analysis at the Harvard Microchemistry Facility, Harvard University. Briefly, gel slices were reduced with DTT and alkylated with iodoacetamide,
35 and then digested with trypsin in 25 mM ammonium bicarbonate buffer. The trypsin digest was analyzed by microcapillary reverse-phase HPLC nano-electrospray tandem mass spectrometry

(μ LC/MS/MS) on a Finnigan LCQ DECA XP Plus quadrupole ion trap mass spectrometer. Preliminary sequencing of peptides was facilitated by database correlation with the algorithm SEQUEST. MS/MS peptide sequences were then reviewed for consensus with known proteins and the results manually confirmed for fidelity. No sequences from the NCBI nr or est databases
5 correlated with this data.

Several attempts to obtain the N-terminal sequence of the undigested protein had failed to generate any sequencing information, suggesting that the N-terminus was blocked. In order to obtain internal sequence information peptides from the trypsin digest of the NuPAGE gel slices containing $\sim 5 \mu\text{g}$ of the desired protein was fractionated by HPLC on a $0.3 \times 150\text{mm}$ C18 column.
10 Three wavelengths were monitored; 205nm (for amide bonds), 277nm and 292nm (for aromatic amino acids Trp and Tyr) via a diode array detector. A few of the best peaks/fractions were screened by MALDI to select peaks for Edman sequencing. Blast database searches using "search for short, nearly exact matches" against the NCBI nr database did not identify any identical sequences with any of the obtained peptide sequences. However, search using a 30 amino acid
15 peptide sequence shown as SEQ ID NO: 12, the longest obtained peptide sequence and confirmed by MS/MS, did identify a candidate putative protein (SEQ ID NO: 2) predicted from the genome sequence of *Streptomyces avermitilis* (GenBank access # BAC74979.1, GI:29610934) that showed weak sequence similarity to SEQ ID NO: 12, the obtained griseoplanus peptide sequence (illustrated in FIG. 10).

20 SEQ ID NO: 12: TVIDVTDFGADPSGKADSAAAVSAAMAHAK

The genome of *Streptomyces griseoplanus* is not available and no related sequences were identified in database searches. Notably, this sequence is not shared by the α -galactosidase described in our prior disclosure (and herein as SEQ ID NO: 1). *Streptomyces avermitilis* and *Streptomyces griseoplanus* are closely related. We therefore tested if *Streptomyces avermitilis*
25 also contained the novel α -galactosidase, since the previous α -galactosidase was demonstrated to be very rare among many of the *Streptomyces* isolates tested (see, U.S.S.N. 10/251,271).

Streptomyces avermitilis (ATCC 31267) was cultured in YM media and the culture supernatant was assayed for secreted α -galactosidase using the AMC labeled B tetrasaccharide and a monosaccharide α -Gal pNP as substrates. As shown in FIG. 11, clear evidence of the
30 presence of α -galactosidase activities in both culture supernatant and pellet lysate as analyzed by B-tetra oligosaccharide. However, the cleavage of a simple substrate α -Gal pNP is negligible by the secreted α -galactosidase activities. In contrast, complete cleavage of α -Gal pNP was observed for the cellular α -galactosidase(s).

The identified putative protein consisted of 625 amino acids (SEQ ID NO: 2) and showed
35 no significant identity to any other known proteins. Back searches with the identified protein sequence identified very few protein sequences with low sequence similarities exclusively from

prokaryotic genomes. All sequences identified were analyzed by multiple sequence analysis as shown in FIG. 12.

EXAMPLE 4: RECOMBINANT EXPRESSION AND CHARACTERIZATION OF IDENTIFIED α -GALACTOSIDASE GENE FROM *STREPTOMYCES AVERMITILIS*

5 The predicted full coding sequence of the identified *Streptomyces avermitilis* gene, 1878 base pairs in length, encoding the putative protein (SEQ ID NO: 2) of 625 amino acids (full length) was amplified by PCR (polymerase chain reaction) using the primer pair AVER1 (5'-
 10 GCGAATTCCCATGGCTCACGGATGCTCCGGAGGG-3' SEQ ID NO: 13)/ AVER3 (5'- GCCTCGAGAAGCTTCTAGTCCGTGACCACGGAGGTGTTC-3' SEQ ID NO: 14), digested with
 NcoI/HindIII restriction enzyme (restriction sites in primers underlined), and cloned into the
 NcoI/HindIII site of the bacterial expression vector pPET28 (Novagen, Cat. No. 70777-3) forming
 the pZQ-B002a construct. Given the fact that the gene possesses an internal NcoI site at position
 1490, insertion of the full-length gene construct was performed using a two-step cloning
 15 procedure. The expression construct was sequenced in full for confirmation. The generated full
 length expression construct pZQ-B002a was used to transform the *E. coli* strain Rosetta (BL21-
 DE3)pLysS (Novagen Cat. No. 70956-3), and plated out on LB-agar plates in the presence of
 Chloramphenicol (34 μ g/ml) and Kanamycin (50 μ g/ml).

For initial analysis of the protein expression, induction was performed at 26°C instead of
 the more common 37°C with a low concentration of inducer (0.1 mM IPTG), a condition favoring
 20 the formation of soluble proteins. The induced cell pellet was lysed by a detergent based chemical
 method and whole lysate was assayed directly under standard condition for the novel B-zyme
 activity without being clarified. As shown in FIG. 13, the cleavage of the AMC labeled blood group
 B tetrasaccharide substrate was readily detectable as indicated by the formation of H trisaccharide
 using crude lysate generated from cultures induced only for 1 hr. This result unambiguously
 25 demonstrates the protein from *Streptomyces avermitilis* (SEQ ID NO: 2) is indeed a novel
 galactosidase, a characteristic activity shared by other members of this family of proteins. FIG. 14
 confirms that SEQ ID NO: 2 can be efficiently expressed in *E. coli*, but is recovered in inclusion
 bodies. Therefore, denaturation, extraction of the enzyme from inclusion bodies, and refolding are
 first necessary, for producing this polypeptide in *E. coli*.

30 EXAMPLE 5: ENZYMATIC CONVERSION OF B RED BLOOD CELLS TO O PHENOTYPE CELLS USING EXPRESSED α -GALACTOSIDASE AS EVALUATED BY ROUTINE TYPING PROTOCOLS

Conversion Protocol One - Enzymatic conversion reactions were performed in 1 ml
 35 reaction mixtures containing 200 mM glycine, pH 6.8, and 3 mM NaCl with 30 % packed red blood
 cells (pRBCs) and enzyme as indicated. Fresh whole blood was obtained from Oklahoma Blood
 Institute (Oklahoma City, OK) and buffy coat removed. RBCs were prewashed 1:1 and 1:4 vol/vol
 in conversion buffer before addition of enzyme, and reactions incubated for 60 min with gentle

mixing at 26°C, followed by four repeat washing cycles with 1:4 vol/vol of saline by centrifugation at 1,000 rpm. The washed enzyme-treated B-ECO RBCs were ABO typed according to standard blood banking techniques using various commercially available monoclonal antibody reagents ((Immucor Gamma Anti-B (Gamma Biologicals/Immucor, Norcross, Ga.); Ortho Anti-B (Ortho Clinical Diagnostics, Raritan, N.J.); and Diagast Anti-B (Diagast Laboratories, France)).

Conversion Protocol Two - B red cells (Beth Israel Deaconess Medical Center, Boston, MA) are drawn into EDTA tubes and stored at 4°C for up to seven days, and are washed three times in PBS (Phosphate Buffered Saline, pH 7.4), and resuspended to 10% in a solution of PBS and 7.5% PEG (pH 7.4). Cells are treated with recombinant α -galactosidase (10-500 U/ml) at 30°C for 180 min while shaking. Cells are washed three times in 0.9% saline and resuspended to 3-5% in saline for typing.

Conversion Protocol Three - B red cells (Beth Israel Deaconess Medical Center, Boston, MA) are drawn into EDTA tubes and leukoreduced B red cells (American Red Cross, New England Region, Dedham, MA) are frozen in Glycerolyte 57, (Baxter Healthcare Corporation, Fenwal Division: Deerfield, IL) according to the AABB Technical Manual, 13th edition, Method 6.6 and stored at -70°C. Prior to enzyme treatment cells are deglycerolized using 9.0% saline, 2.5% saline, and 0.9% saline (see, Method 125 of Immunohematology Methods by the American Red Cross), then resuspended to a hematocrit of 50% in a solution of PBS and 7.5% PEG (pH 7.4) and recombinant α -galactosidase (200 U/ml) is added. Reactions are incubated at 37°C with shaking for 4 hours, followed by three washes in 0.9% saline, and final suspension to 3-5% in saline for typing.

Conversion Protocol Four - Origin and storage of cells is the same as described under protocol B. Deglycerolized red cells are washed twice in PCI (pH 7.4) with 150 mM NaCl and resuspended to a hematocrit of 50% in PCI (pH 7.4) with 150 mM NaCl. Cells are treated with recombinant α -galactosidase (200 U/ml) at 37°C with shaking for 4 hours, followed by three washes in 0.9% saline, and final suspension to 3-5% in saline for typing.

Approved typing reagents used in hemagglutination assays are murine monoclonal antibodies and plant lectins obtained from Ortho Clinical Diagnostics, Raritan, N.J.; Gamma Biologicals/Immucor, Norcross, Ga. Non-FDA approved reagents included murine monoclonal anti-B antibodies to blood group B variants produced by H. Clausen (Clausen *et al.*, Proc. Natl. Acad. Sci. USA 82(4): 1199-203, 1985, Clausen *et al.*, J Biol Chem. 261(3): 1380-7, 1986, Clausen *et al.*, Biochemistry 25(22): 7075-85, 1986, Clausen *et al.*, J Biol Chem. 262(29): 14228-34, 1987). Typing reagents are used according to the manufacturers recommendations and other monoclonal antibodies as determined by titrations.

Hemagglutination Assay (room temperature).

A 3-5% suspension of washed red cells in isotonic blood bank saline is prepared. One drop (approx 50 microliters) of anti-B antibody reagent is added. One drop (approx 50 microliters) of the red cell suspension is added. Tubes are mixed and centrifuged for 15 seconds at 3500 rpm. Cells are resuspended by gentle agitation and examined macroscopically for agglutination. The agglutination is graded according to Method 1.8 in the AABB Technical Manual, 13th edition.

As described in the previous examples, preferred enzymes for use in removing blood group B epitopes from red cells are likely to have particularly good kinetic properties with oligosaccharide substrates resembling the blood group B antigens. Such preferred kinetic properties could be represented by preferred or exclusive substrate specificities for the blood group B oligosaccharides, and low or no activity with simple monosaccharide derivatives such as monosaccharide-pNP substrates. Preferred kinetic properties could also be represented by a particularly low Km for relevant substrates. Further preferred kinetic properties consist of neutral pH optimum of reactions with relevant blood group active substrates, and other reaction conditions that are compatible with the integrity and functions of red cells. Other preferred properties of the enzyme such as size, charge, solubility, and other physico-chemical properties may also relate to performance in enzymatic conversion of red cells. The novel α -galactosidase with improved kinetic properties was identified from various bacterial strains as described and provides an enzyme with the above mentioned preferred characteristics, that exhibits superior performance in red cell conversions.

Table 3A: Agglutination Results of human red cells converted with FragA or FragB recombinant α galactosidases.

Routine Conversion Protocol (200 Glycine, pH 6.8, 3 mM NaCl)		Immucor Anti-B		Diagast Anti-B	
Frag B enzyme	Dose μ g/ml	IS	4°C	IS	4°C
Human B Cells	10	0	0	0	0
	5	0	0	0	0
	2.5	0	0	0	0
	1.25	0	0	0	0
	0.625	0	W+	W+	1+
	0.3125	0	1+	1+	1+
Frag A enzyme					
Human B Cells	10	0	0	0	0
	5	0	0	0	0
	2.5	0	0	0	0
	1.25	0	0	0	0
	0.625	0	W+	W+	1+
	0.3125	1+	1+	1+	2+

EXAMPLE 6: CLONING AND DNA SEQUENCING OF α -GALACTOSIDASE FROM *STREPTOMYCES GRISEOPLANUS* STRAIN #2357 AND DEDUCTION OF ITS AMINO ACID SEQUENCE

The isolation and purification of the endogenous α -galactosidase from *S. griseoplanus* 2357 was described in Example 2. The partial amino acid sequencing of purified α -galactosidase

that generated a 30 amino acid peptide was described in Example 3. Blast search using this peptide against 'nr' database (GenBank) identified a family of putative α -galactosidases. The sequences for the 5 α -galactosidases were submitted to the EMBL/GenBank/DDBJ databases and assigned the following accession numbers: AM109953 (*Streptomyces avermitilis*), AM109954 and AM109955 (*Bacteroides fragilis*), AM109956 and AM109957 (*Bacteriodes thetaiotaomicron*). Multiple sequence alignment of putative α -galactosidases identified a few conserved regions (FIG. 15). The open reading frame encoding a putative α -galactosidase from *Streptomyces griseoplanus* was cloned based on 5' and 3' rapid amplification of genomic ends (RAGE). Initial degenerate primers were based on the conserved regions determined from multiple sequence alignment of putative α -galactosidase sequences. Degenerate sense and anti-sense primers dAVER7 (5'-TTCGGXGTXTXKXKGCAGTWCAGXGAGAA-3' SEQ ID NO: 15)/dAVER9 (5'-GTXCCXTGXATXTTXATXGGXTCXTCGTG-3' SEQ ID NO: 16), where X= inosine and K= G or T, were used to PCR amplify a single 185bp BZyme specific DNA fragment from *Streptomyces griseoplanus* genomic DNA. PCR product was cloned into pCR4 vector (Invitrogen) and sequenced generating pCR4-dAVER7/9. *Streptomyces griseoplanus* α -galactosidase specific primers GRIS10 (5'-ATCGACTCGGTACCTTCAAGGCCGAC-3' SEQ ID NO: 17) and GRIS11 (5'-AAGACGCTGTTGGTGATGCGTACGGTGC-3' SEQ ID NO: 18) were derived from pCR4-dAVER7/9. *Streptomyces griseoplanus* genomic DNA was endonuclease treated to completion with restriction endonuclease HaeII, size fractionated by 0.8% agarose gel electrophoresis and 2-3kbp fractionated DNA was purified by Qiagen purification (Qiagen). Purified DNA was ligated to a double stranded HaeII adapter EBRETTE3 (5'-GCGCTCGAAATTAACCCTCACTAAAGGGGAATTCGGTACCCTCGAGGCGC-3' SEQ ID NO: 19)/EBRETTE4 (5'-CTCGAGGGTACCGAATTCGGAA-3' SEQ ID NO: 20) encoding a T7 binding site (underlined) and HaeII restriction overhang (shown in italics). Adapter ligated DNA was used in 5' RAGE using 10ng adapter ligated DNA, T7/GRIS11 or 3'RAGE using 10ng T7/GRIS10. Generated 5' and 3' RAGE products were cloned into pCR4 generating 5'-T7/GRIS11-pCR4 and 3'-T7/GRIS10-pCR4 and fully sequenced. The overlapping 5'-T7/GRIS11-pCR4 and 3'-T7/GRIS10-pCR4 sequences represent 1593bp of the full coding BZyme gene sequence. Remaining 5' and 3' sequence was obtained by repeated RAGE on fractionated BamHI digested *Streptomyces griseoplanus* genomic DNA, BamHI adapter EBRETTE3/6 (5'-GATCGCGCCTCGAGGGTACCGAATTCGGAA-3' SEQ ID NO: 21) ligated (BamHI overhang shown in italics). Complete 5' sequence was obtained using 10ng adapter ligated DNA and T7/GRIS22 (5'-CGCTTCGGCGTCCGTTCCGGGCCAG-3' SEQ ID NO: 22) and 3' sequence using T7/GRIS24 (5'-CCGGTGCACCGCAACGTCCTCATC-3' SEQ ID NO: 23). Generated 5' and 3' RAGE products were cloned into pCR4 generating 5'-T7/GRIS22-pCR4 and 3'-T7/GRIS24-pCR4 and fully sequenced. 5'-T7/GRIS22-pCR4 contained a predicted initiating start methionine and 3'-T7/GRIS24-pCR4 contained an in frame stop codon, completing the full 2184 bp coding sequence of *Streptomyces griseoplanus* α -galactosidase gene (SEQ ID NO: 24) encoding a 727 amino acid α -galactosidase (SEQ ID NO: 25). The sequence was submitted to GenBank (accession number

AM259273). The regions in protein sequence of the α -galactosidase from which the primers were derived are described in FIG. 16.

EXAMPLE 7: RECOMBINANT EXPRESSION AND CHARACTERIZATION OF IDENTIFIED α -GALACTOSIDASE GENE FROM BACTEROIDES FRAGILIS

5 FragB α -galactosidase (SEQ ID NO: 6) expression construct was cloned from genomic bacterial DNA by PCR. The FragB α -galactosidase gene, lacking the coding region for the putative amino terminal signal peptide 1-24, was lifted from *Bacteroides fragilis* genomic DNA (ATCC 25285D) by PCR using primers BFRAGB2 (5'-
 GCGGGATCCCGGGATGGGACGTGTTTATGACATTTCCAGTTTGGC-3' SEQ ID
 10 NO: 24)/BFRAGB3 (5'-GCCTCGAGAAGCTTTTCACTCTGAAATCTTCACGTTTGTCACTCG-3' SEQ ID NO: 25) and amplified using Pfu Ultra polymerase (Stratagene). Restriction enzyme overhangs for BamH I and Hind III in the above primers are underlined. After digestion with BamH I and Hind III, the amplified polynucleotide products were inserted into the bacterial expression vector pET28 (Novagen) in frame and downstream of the plasmid encoded 6xHis tag to generate
 15 plasmid pZQ-B006a. For the construction of a non tagged expression vector, the 6xHis tag in pET28 vector was removed by NcoI/BamHI digestion followed by insertion of a double stranded oligo PETNCBAF (5'-CATGGATCCCAGGCCTCCGGATG-3' SEQ ID
 NO: 26)/(GATCCATCCGGAGGCCTGGGATC-3' SEQ ID NO: 27) creating plasmid pET28- δ His. The FragB construct described above encoding the His-tagged protein was sub-cloned into the
 20 pET28- δ His BamHI/HindIII site to create plasmid pZQ-B006c for the expression of the untagged FragB α -galactosidase. All constructs were fully sequenced on a 377 ABI Prism instrument (Applied Biosystems). For protein expression, pZQ-B006c was transformed into *E. coli*, Rosetta2 (DE3) (Invitrogen). The *E. coli* clone was grown in 1X Terrific Broth (Sigma), supplemented with 34 μ g/mL of Chloramphenicol and 50 μ g/mL of Kanamycin at 37 °C, 220 rpm to ~0.6 OD at 600
 25 nm and IPTG was added to 0.5 mM to induce the target protein expression. The culture was harvested after 3 hrs, by centrifugation at 3000 xg for 30 min. The cell pellet was stored at -20°C. The cell pellet harvested from 350ml culture was lysed using 1X BugBuster (Invitrogen) in 25 mM NaOAc, pH 5.5/10 mM NaCl, supplemented with 5 μ L of Benzonase (Invitrogen), and stirred for 1 hr at room temperature. The whole lysate was clarified by centrifugation at 40,000 x g, 5°C, for 30
 30 min. The cell debris, containing over 90% enzyme activity, was re-suspended in 10 mM NaPO₄, pH 6.8/400 mM NaCl and stirred at room temperature for 30 min. High speed centrifugation was repeated to recover the supernatant containing the enzyme activity. The resulting supernatant was loaded onto a 5-ml Hydroxyapatite column, pre-equilibrated with 10 mM NaPO₄, pH 7.0. The column was washed with 20 mL of equilibration buffer, followed by elution using a NaPO₄
 35 gradient, pH 7.0, from 10 to 400 mM. The enzyme activities, eluted between 200-400 mM NaPO₄, pH 7.0, were pooled, concentrated and buffer exchanged with an Amicon (Grace) centrifugal device Plus 70, into 10 ml of 40 mM Tris, 400 mM NaCl/pH 7.5. The protein solution was allowed

to pass through a 5-ml Phenyl Sepharose High Performance column (Amersham), pre-equilibrated with the dialysis buffer, and the column was washed with 10 mL of dialysis buffer after loading. The flow through and wash were pooled, adjusted to pH 8.5 with 1 M Tris, diluted with equal volume of H₂O. The resulting protein solution was subjected to another passage column step, 2.3
5 mL of Macro-Prep High Q, pre-equilibrated with 40 mM Tris, pH 8.5/10 mM NaCl, and the column was washed with 10 mL of equilibration buffer. The flow through and wash were pooled and buffer exchanged into 7 ml of 10 mM NaPO₄, pH 6.8/50 mM NaCl. The protein concentration was determined by Pierce's BCA Protein Assay Kit.

The purified FragB α -galactosidase was evaluated for its ability to cleave the branched carbohydrate chain substrate B-tetra under standard conditions (1 nmole of substrate in 10 μ L of
10 100 mM NaPO₄, pH 6.8/50 mM NaCl), and the purified enzyme demonstrated extremely high specific activity toward the B-tetra substrate: ~5-10 U/mg. The pH optimum of the purified FragB α -galactosidase was evaluated with the B-tetra-AMC substrate across a pH range of 2.0 to 9.0 and the results are shown in FIG. 17. The FragB enzyme has a broad optimal pH range between 4.5
15 and 7.5. Analysis with the more sensitive/quantitative colorimetric assay gives the similar conclusion as shown in FIG. 18, although the enzyme's activity was seen at the low end of the pH range tested, *i.e.*, activity was observed down to ~pH 4.2, a subtle difference in activity which is not detectable by using the TLC based AMC-B-tetra assay. Therefore, the novel enzyme can be used successfully under acidic conditions to neutral condition and even slightly basic conditions,
20 *i.e.*, about pH 4 to about pH 7.4 or greater. In currently preferred embodiments, the enzyme is used within a pH range of about 6.0 to about 7.5, and more preferably about pH 6.5 to about pH 7.5. The currently most preferred pH range for the novel enzyme are those conditions mimicking the physiological pH of circulating arterial or venous blood, in order to minimize pH effects on the blood cells themselves and not because of pH limitations on enzyme activity and performance.

The FragA α -galactosidase (SEQ ID NO: 5) was cloned similarly by PCR from the same genomic DNA as for FragB α -galactosidase, and expressed similarly as His₆ tag protein at the N-terminus in Rosetta (DE3) pLysS (Novagen). Expressed soluble protein was purified to
homogeneity by successive immobilized metal affinity chromatography (IMAC), cation and anion exchange chromatography. The purified protein was shown to be similar to the endogenous *S.*
30 *griseoplanus* α -galactosidase (strain 2357) in terms of specific activity for branched substrate, substrate specificity and pH optima. Analysis of activity with a blood group B tetrasaccharide-AMC substrate in the pH range of 2-9 showed that the enzyme has a broad optimum between 5 and 7.5. The substrate specificity of the FragA α -galactosidase was determined with a diverse panel of oligosaccharide structures and a remarkably stringent specificity for α 1-3 linked galactose in the
35 branched blood group B structure was found (FIG. 20). The enzyme cleaved neither α 4Gal linkages found in P¹ and P^k blood group antigens nor the α 3Gal linkage in linear B structure without fucose.

EXAMPLE 8: BACTEROIDES FRAGILIS α -GALACTOSIDASE EFFICIENTLY CLEAVES LINEAR B OLIGOSACCHARIDES AT A NEUTRAL PH

Further analysis of the substrate specificity of recombinant purified FragB α -galactosidase surprisingly revealed that this enzyme (in contrast to the earlier purified α -galactosidase from *S. griseoplanus*) exhibited low activity with the Gal α -pNP substrate (~1.6 U/ml using the buffer system 100 mM NaPO₄, pH 6.8/50 mM NaCl) (Table IV).

Table IV. Comparison of the specific activities (U/mg) of FragB α -Galactosidase and Coffee Bean α -Galactosidase.

Substrate	Coffee Bean	<i>B. fragilis</i> (FragB) ¹
Gal α -pNP	32 (pH 6.5) ²	1.6 (pH 6.8)
Gal α 1-3(Fuc α 1-2)Gal β 1-4Glc-AMC	0.017 (pH 5.5) ⁴	9.4 (pH 6.8)

¹This work.

²Derived from Zhu, A., Monahan, C., Zhang, Z., Hurst, R., Leng, L. & Goldstein, J. (1995) Arch Biochem Biophys **324**, 65-70.

³not determined.

⁴Derived from US patent application, publication number 20050208655.

This prompted us to test the substrate specificity with substrates having different α 1-3Gal and α 1-4Gal linkages. As shown in FIG. 19, FragB showed high activity with linear α 1-3Gal linkages (B-di and Linear B) in addition to the blood group B oligosaccharide structures. Interestingly, the activity of FragB with the Gal α 1-3Gal disaccharide was very high (~12 U/mg at pH 6.8) suggesting that this enzyme is suitable for efficient cleavage of the linear B antigen (Gal α 1-3Gal β 1-4GlcNAc β 1-R, where R is any oligosaccharide structure) also known as the Galili antigen (Galili U (2005) Immunol Cell Biol 83:674-86). The Galili antigen is a major xenotransplantation barrier antigen found on most animal tissues except old world monkeys and man (Galili U (2005) Immunol Cell Biol 83:674-86). Xenotransplantation of *e.g.* pig tissues and cells into man results in hyperacute rejection mainly due to presence of high titers in man of IgG antibodies to the Galili antigen. To date only broadly reactive α -galactosidases such as Coffee bean derived enzymes with acidic pH optimums have been used for cleavage of the Galili antigen from cells and tissues. This may constitute a significant problem since all animal cells express large quantities of *e.g.* the globoseries P^k glycolipid structure (Gal α 1-4Gal β 1-4Glc β 1-Ceramide). It is more desirable to use α -galactosidase enzymes with higher efficiency, specificity and neutral pH optimum, such as those disclosed herein.

The FragB α -galactosidase was specific for α 1-3Gal linkages, and no significant cleavage of several α 1-4Gal linkages (P₁ and P^k) was observed. This is similar to the α -galactosidase obtained from *S. griseoplanus*, but represents a property that is different from any other known α -galactosidase including the enzyme derived from Coffee bean. The results are summarized in Table V.

Table V. Substrate specificity of the α -galactosidases.

Substrates	Blood Group Specificity	Coffee Bean ¹	<i>S. griseoplanus</i> ¹	<i>B. fragilis</i> ² (FragB)
Gal α -pNP	NA ³	+ ⁴	- ⁵	+
Gal α 1-3Gal	B (B-di)	+	-	+
Gal α 1-3Gal β 1-4GlcNAc	Linear B	+	-	+
Gal α 1-3(Fuca1-2)Gal	B-tri	+	+	+
Gal α 1-4Gal	P ₁	+	-	-
Gal α 1-4Gal β 1-4Glc	P ^k	+	-	-
GalNAc α 1-3(Fuca1-2)Gal	A-tri	-	-	-
Gal α 1-3(Fuca1-2)Gal β 1-4Glc-AMC	B-tetra	+	+	+
GalNAc α 1-3(Fuca1-2)Gal β 1-4Glc-AMC	A-tetra	-	-	-

¹Derived from US patent 20050208655 (Ref.) and Zhu, A., Monahan, C., Zhang, Z., Hurst, R., Leng, L. & Goldstein, J. (1995) Arch Biochem Biophys **324**, 65-70.

5 ²This work.

³Not applicable.

⁴Activity readily detectable under assay conditions.

⁵Activity not readily detectable under assay conditions.

The surprising finding that the FragB α -galactosidase may represent a novel highly efficient enzyme for cleavage of linear B (Galili epitopes) prompted us to test the suitability and efficiency of this enzyme in removing such epitopes from cell surfaces. Rabbit red blood cells contain glycolipids and glycoproteins oligosaccharide chains similar to human red cells, but while oligosaccharide chains in human red cells terminate in ABH structures depending on blood group status, oligosaccharides of rabbit red cells terminate with linear B (Galili epitopes) structures (Gal β 1-3Gal β 1-4GlcNAc). The lectin Bandeiraea (Griffonia) simplicifolia IB4 is generally used to detect linear B (Galili epitopes) structures (Galili U (2005) Immunol Cell Biol 83:674-86), and as shown in Table VI this lectin strongly agglutinates rabbit (but not human) red cells.

Table VI: Agglutination of human and rabbit red cells with IB4 lectin and Routine Monoclonal Anti-B Typing Reagents.

RBC Type	Immucor/Gamma Anti-B		Diagast Anti-B		B4 Lectin 10ug/ml
	IS	4°C	IS	4°C	
Native Rabbit Cells	2+	4+	4+	4+	4+
Native Human B Cells	4+	4+	4+	4+	1+
Native Human A Cells	0	0	0	0	0
Native Human O Cells	0	0	0	0	0

20

Rabbit red cells therefore serve as an excellent model for analysis of the efficiency of α -galactosidases in the removal of the immunodominant α 1-3Gal residue from linear B (Galili epitopes) structures from cell surfaces. Since we have previously developed an efficient conversion process for removal of immunodominant A-type antigens from blood cells using a purified *C. meningosepticum* α -N-acetylgalactosaminidase enzyme, operative using a Glycine pH 6.8 buffer system, we tested the same conditions for FragB cleavage of rabbit red cells in

25

comparison with human blood group B red cells. As shown in Table VII, FragB efficiently removed IB4 lectin agglutination of rabbit red cells at very low doses, almost comparable to what was required for FragB cleavage of blood group B from human red cells. The agglutination of rabbit red cells by the IB4 lectin was almost completely abolished by the enzyme treatment with 10g/ml enzyme dose giving only a microscopic reading (M+). Higher concentrations of enzyme or longer incubation complete abolish reactivity. The homologous gene FragA, however, only cleaved blood group B from human red cells.

Table VII: Agglutination Results of human and rabbit red cells digested with FragA or FragB glycan modifying enzymes.

Routine Conversion		Immucor Anti-B		Diagast Anti-B		B4 Lectin
Frag B enzyme	Dose ug/ml	IS	4°C	IS	4°C	10ug/ml
Human B Cells	10	0	0	0	0	0
	5	0	0	0	0	0
	2.5	0	0	0	0	0
	1.25	0	0	0	0	0
	0.625	0	W+	W+	1+	0
	0.3125	0	1+	1+	1+	0
Rabbit Cells	10	0	0	0	W+	M+
	5	0	0	0	W+	Vw+
	2.5	0	0	0	W+	W+
	1.25	0	0	0	1+	1+
	0.625	0	W+	W+	3+	1+
	0.3125	0	2+	4+	4+	2+
Frag A enzyme						
Human B Cells	10	0	0	0	0	0
	5	0	0	0	0	0
	2.5	0	0	0	0	0
	1.25	0	0	0	0	0
	0.625	0	W+	W+	1+	0
	0.3125	1+	1+	1+	2+	0
Rabbit Cells	10	4+	4+	4+	4+	3+
	5	4+	4+	4+	4+	3+
	2.5	4+	4+	4+	4+	3+
	1.25	4+	4+	4+	4+	3+
	0.625	4+	4+	4+	4+	3+
	0.3125	4+	4+	4+	4+	3+

10 This demonstrates that the purified FragB polypeptide isoform described as part of the novel α -galactosidase gene family reported herein, has unique substrate specificities that are different from several other members within this family, including the related genes observed in Streptomyces. Furthermore, that the FragB polypeptide described is suitable for enzymatic removal of immunodominant blood group B antigens from red cells, as well as for removal of the xenotransplantation Galili antigen from blood cells. The purified FragB polypeptide is thus superior to other currently known enzymes, with respect to its pH optimum (most preferably pH 6.5 to pH 7.5), its restricted substrate specificity to Gal α 1-3 linkages and its observed high specific activity.

15 Enzymatic removal of the immunodominant α 1-3 linked terminal Galactose of the Galili antigens has important applications in the xenotransplantation field. The Galili antigen constitutes

the most important barrier for xenotransplantation of organs, tissues, tendons, ligaments and cells from animals to man, and is the primary cause of the hyper-acute rejection phenomenon (Galili U (2005) Immunol Cell Biol 83:674-86). Approximately 10 % of normal healthy individuals serum IgG is directed to the Galili antigen. Enzymatic removal of the terminal galactose residue will expose
5 common structures found in man, which may abrogate hyperacute rejection (Galili U (2005) Immunol Cell Biol 83:674-86).

The process described for enzymatic removal of the Galili antigen on rabbit cells in which cells are washed and incubated with the FragB α -galactosidase in a suitable buffer at neutral pH for a period of time results in efficient removal of the Galili epitope (Table VII). A similar process
10 applied to animal organs, tissue, tendons, ligaments and cells, results in efficient removal of exposed Galili antigens.

The preferred process involves contacting the animal tissues or cells with the FragB α -galactosidase (or homologous members of the gene family with similar enzymatic activities) in a suitable buffer such as physiological saline, Glycine or other similar buffer systems described
15 herein, at neutral pH of 5.5 to 8.0 and more preferably 6.5 to 7.5. The enzyme dose and time required for enzymatic removal of the immunodominant α 1-3 linked terminal Galactose generally follows the digestion parameters described above for blood cells, but is evaluated empirically, as is determined by lectin and antibody based immunoassays such as immunocytology, immunohistology and ELISA using such suitable lectins such as the IB4 lectin or suitable
20 monoclonal antibodies reactive with the Galili epitope (Galili U (2005) Immunol Cell Biol 83:674-86). When the reaction is complete, the enzyme modified animal organ, tissue, tendon, ligament or cells are washed with an appropriate buffer solution (such as physiological saline) to remove the enzyme solution. The animal tissues or cells lack immunodominant Galili antigens, and can now be used as an appropriate xenotransplant into a human subject in need of such a transplant. An
25 example of this is an antigenically modified porcine ligament, which is used for the reconstruction of ruptured anterior cruciate ligament in a human patient. See for example, United States Patent 6,402,783.

Prior to treatment, the outer surface of the xenograft may optionally be pierced to increase permeability to agents used to render the xenograft substantially non-immunogenic. A sterile
30 surgical needle such as an 18 gauge needle may be used to perform this piercing step, or, alternatively a comb-like apparatus containing a plurality of needles may be used. The piercing may be performed with various patterns, and with various pierce-to-pierce spacings, in order to establish a desired access to the interior of the xenograft. Piercing may also be performed with a laser. In one embodiment of the invention, one or more straight lines of punctures about three
35 millimeters apart are established circumferentially in the outer surface of the xenograft.

Prior to implantation, the ligament xenograft of the invention may be treated with limited digestion by proteolytic enzymes such as ficin or trypsin to increase tissue flexibility or coated with anticalcification agents, antithrombotic coatings, antibiotics, growth factors, or other drugs which

may enhance the incorporation of the xenograft into the recipient knee joint. The ligament xenograft of the invention may be further sterilized using known methods, for example, with additional glutaraldehyde or formaldehyde treatment, ethylene oxide sterilization, propylene oxide sterilization, or the like. The xenograft may be stored frozen until required for use.

5 The ligament xenograft of the invention, or a segment thereof, may be implanted into a damaged human knee joint by those of skill in the art using known arthroscopic surgical techniques. Specific instruments for performing arthroscopic techniques are known to those of skill in the art, which ensure accurate and reproducible placement of ligament implants. Initially, complete diagnostic arthroscopy of the knee joint is accomplished using known methods. The
10 irreparably damaged ligament is removed with a surgical shaver. The anatomic insertion sites for the ligament are identified and drilled to accommodate a bone plug. The size of the bone plug can be about 9-10 mm in width by about 9-10 mm in depth by about 20-40 mm in length. The xenogeneic ligament is brought through the drill holes and affixed with interference screws. Routine closure is performed.

15 Using the polypeptides of the present invention thus permits removal of the Galilli antigen from many different tissues types, using the modification procedures described herein and as may be further adapted to the particular tissues in view of the teachings provided, by a skilled artisan. These modified tissues are used for a variety of transplant procedures where non-immunogenic xenotransplants are required, as is described in the following: to create *e.g.* substantially non-
20 immunogenic injectable collagen (*see*, U.S. Patent 7,064,187); for bone xenografts (*see*, U.S. Patent 6,972,041); for soft tissue and proteoglycan-reduced soft tissue xenografts (*see*, U.S. Patent 6,758,865 and 6,455,309); xenograft heart valves (*see*, U.S. Patent 6,383,732); and meniscal xenografts (*see*, U.S. Patent 6,093,204 and 5,984,858).

In another particularly preferred embodiment, the invention provides for tissue matrices preferably
25 those made from α 1,3-galactose-deficient tissues (*see*, U.S. Patent 6,933,326 and U.S. Patent Application 20050159822 and 20050028228). Methods of making and using these tissue matrices are described in the above patent and applications, and the de-galactosylation of the tissues is accomplished using the novel α 3 galactosidases as described herein (SEQ ID NO: 2-9, or active fragments or functional equivalents thereof).

Claims

We claim:

1. A modified red blood cell comprising: a group B erythrocyte or a group AB erythrocyte that lacks immunodominant B epitopes, as determined by serological typing.
- 5 2. The modified red blood cell as described in claim 1, wherein the blood cell lacks immunodominant B epitopes but retains either α 1,3 Gal epitopes or α 1,4 Gal epitopes.
3. A modified red blood cell prepared by the method comprising: obtaining a group B erythrocyte or a group AB erythrocyte, suspending the erythrocyte in a buffer solution having a neutral pH, and contacting the erythrocyte with a polypeptide having α 3 galactosidase activity, thereby substantially cleaving from the erythrocyte the
10 immunodominant B epitopes but not but the α 1,3 Gal epitopes or α 1,4 Gal epitopes.
4. The modified red blood cell of claim 3, wherein the enzyme used to process the erythrocyte includes the polypeptide or the nucleotide sequence encoding same specified as SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID
15 NO: 7, SEQ ID NO: 8 or SEQ ID NO: 9.
5. The modified red blood cell of claim 4, wherein the enzymatic processing of the erythrocyte is performed at from about pH 6.0 to 8.0.
6. The modified red blood cell of claim 5, wherein the enzymatic processing of the erythrocyte is performed at from about pH 6.5 to 7.5.
- 20 7. The modified red blood cell of claim 5, wherein the enzymatic processing of the erythrocyte is performed at from about pH 7.0 to 7.5.
8. The modified red blood cell of claim 5, wherein the enzymatic processing of the erythrocyte is performed using 1-10 micrograms of enzyme per ml blood cells.
9. A method of modifying a red blood cell, comprising: obtaining a group B or group AB
25 erythrocyte, suspending it in a buffer solution having a neutral pH, and contacting it a polypeptide having the sequence specified as: SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8 or SEQ ID NO: 9, thereby cleaving the branched immunodominant B epitopes on the group B or group AB erythrocyte as determined by serological typing, or hemagglutination assays.
- 30 10. A method of treating a subject, comprising: identifying a human subject in need of blood, the subject being seropositive for anti-B antibodies; obtaining a modified blood cell preparation of claim 1, or a blood cell preparation obtained by the methods of claim 3, and; transfusing the blood cell preparation into the subject, wherein the subject does not immunologically reject the transfused blood cells.

11. A purified enzyme, comprising: a polypeptide including the sequence specified by SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8 or SEQ ID NO: 9, wherein the enzyme displays alpha galactosidase branched substrate specificity and a neutral pH optima.
- 5 12. A purified enzyme, comprising:
- (a) a polypeptide having at least 10 amino acids in the following sequence numbered accordingly when aligned with SEQ ID. NO:2:
- 10 M at residue 10; G at residue 47; G at residue 84; Y at residue 86;
Y at residue 99; N at residue 102; K at residue 114; T at residue 127;
G at residue 130; G at residue 132; G at residue 139; N at residue 156;
D at residue 160; P at residue 164; G at residue 205; R at residue 277;
R at residue 281; F at residue 287; G at residue 308; Q at residue 312;
15 I at residue 317; R at residue 333; D at residue 340; G at residue 346;
G at residue 349; G at residue 360; D at residue 363; D at residue 364;
N at residue 367; H at residue 369; G at residue 370; T at residue 371;
G at residue 396; E at residue 462; N at residue 463; T at residue 465;
T at residue 467; P at residue 468; R at residue 483; G at residue 484;
L at residue 486; T at residue 489; N at residue 498; I at residue 508;
20 D at residue 513; W at residue 517; E at residue 519; G at residue 521;
D at residue 525; I at residue 528; N at residue 531; F at residue 533;
I at residue 549; P at residue 553; I at residue 573; A at residue 590;
G at residue 595; N at residue 601; and, I at residue 629;
- (b) wherein the polypeptide has at least 20% identity with SEQ ID NO: 2, and wherein the polypeptide has α 3 galactosidase activity.
- 25 13. A purified enzyme, comprising a polypeptide including nine contiguous amino acids having the sequence DD(P/A)(V/I)N(V/I)HGT, wherein the polypeptide has α 3 galactosidase activity.
14. A purified enzyme, comprising a polypeptide including twenty-one contiguous amino acids having the sequence: DXXXW(Y/F)E(S/T)GXXXD(L/V)(L/T)I(K/R)XNXF, where X can be
30 any amino acid, wherein the polypeptide has α 3 galactosidase activity.
15. The purified enzyme of claims 11, 12, 13, or 14 further comprising functional equivalents thereof.
16. The purified enzyme of claim 15, wherein the enzyme includes a truncated variant lacking a signal sequence.
- 35 17. The modified red blood cell of claim 3, wherein the erythrocyte is enzymatically processed to remove the immunodominant B epitopes using the alpha galactosidase enzyme of claims 12, 13, 14, 15, or 16.
18. A method of producing a recombinant enzyme, comprising: obtaining a nucleic acid encoding SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6,
40 SEQ ID NO: 7, or SEQ ID NO: 8; expressing the nucleic acid in a cell transfected

- therewith; inducing expression of the nucleic acid encoding the enzyme; and purifying the expressed enzyme from the cell, wherein the enzyme has α 3 galactosidase activity.
19. A non-naturally occurring prokaryotic cell, comprising: an expression vector not found in a wild-type prokaryotic cell, the expression vector having a nucleic acid sequence encoding a polypeptide including the sequence specified by SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 8 or SEQ ID NO: 9, and wherein the polypeptide has α 3 galactosidase activity.
20. The purified prokaryotic α -galactosidase of claim 11 or claim 12, having restricted specificity for cleaving Gal α 1-3 linkages, but not cleaving Gal α 1-4 linkages.
21. The purified prokaryotic α -galactosidase of claim 20 having specificity for cleaving branched Gal α 1-3 linkages such as in blood group B structures.
22. The purified prokaryotic α -galactosidase of claim 20 having specificity for cleaving linear and branched Gal α 1-3 linkages, such as in linear B and blood group B structures.
23. The purified prokaryotic α -galactosidase of claims 11, 12, 20 and 22, wherein the α -galactosidase cleaves linear and branched Gal α 1-3 linkages over a pH range of 5.0 to 7.5.
24. A method of preparing a tissue for xenotransplantation, comprising obtaining a tissue from a non-human animal source, incubating the tissue with a polypeptide having α 3-galactosidase activity thereby removing immunodominant α 1-3 linked terminal galactose residues from the tissue, and isolating the tissue from the polypeptide and the enzymatically removed galactose, thereby rendering the tissue suitable for xenotransplantation into humans.
25. The method of claim 24, wherein the tissue is porcine connective tissue.
26. The method of claim 25, wherein the porcine connective tissue is a ligament.
27. The method of claim 24, wherein the tissue is an organ including liver, kidney, or heart.
28. The method of claim 24, wherein the tissue is non-immunogenic injectable collagen; bone xenografts; soft tissue and proteoglycan-reduced soft tissue xenografts; xenograft heart valves; meniscal xenografts; and tissue matrices, wherein the tissues are α 1,3-galactose-deficient tissues that have been modified using an α 3 galactosidase of any of SEQ ID NO: 2-9.

FIG. 1.

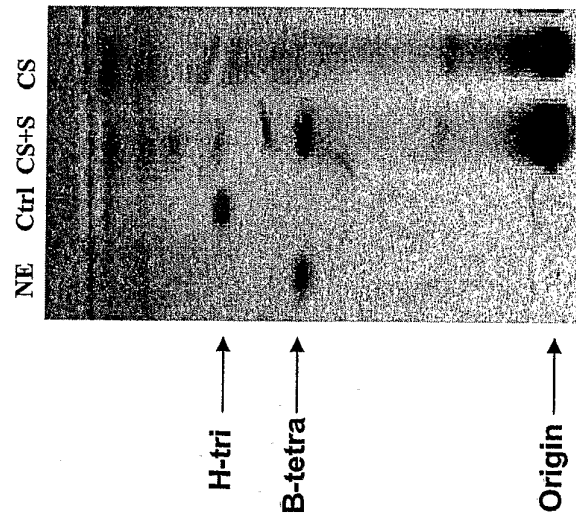


FIG. 2.

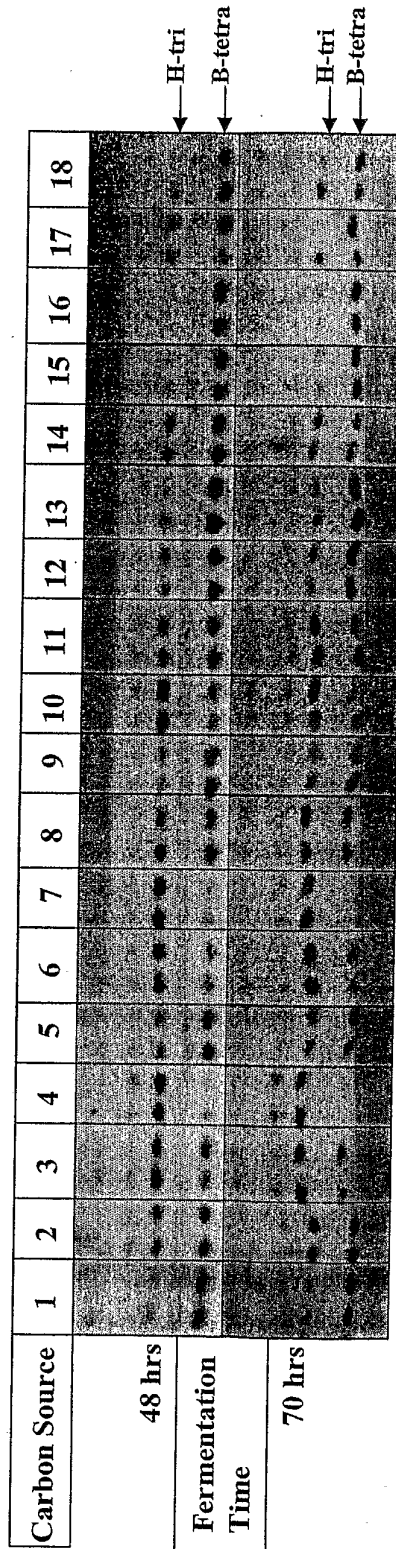
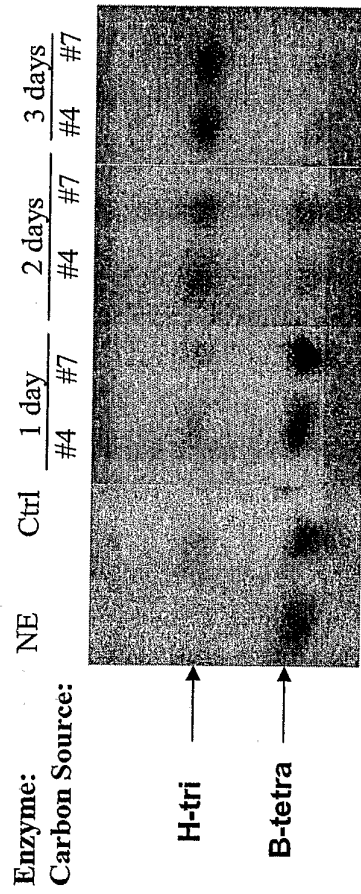


FIG. 3.



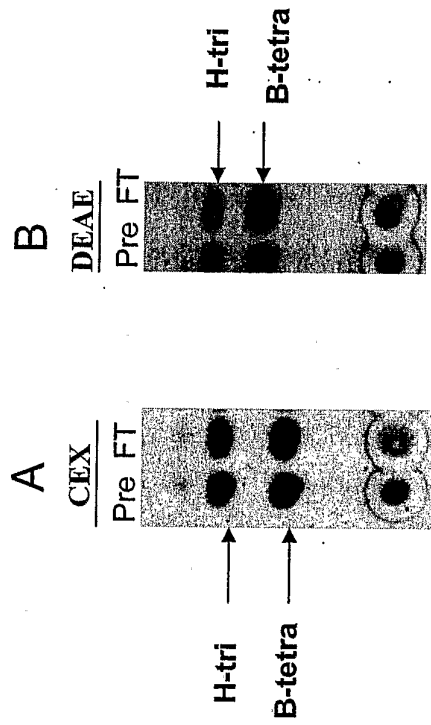


FIG. 4.

FIG. 5.

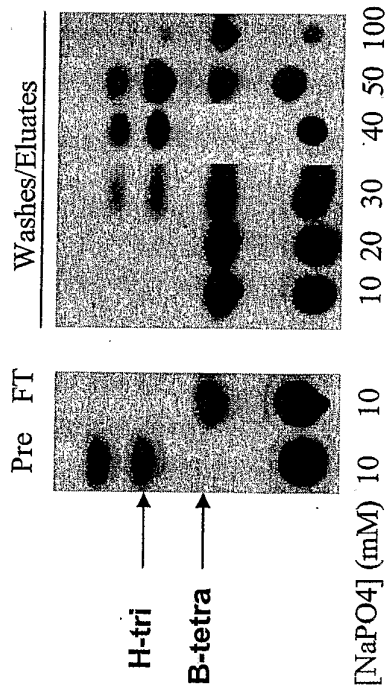


FIG. 6.

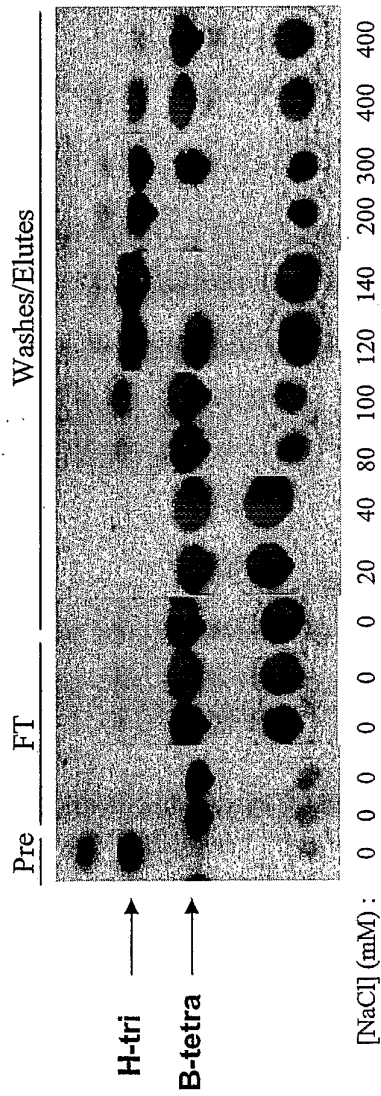


FIG. 7.

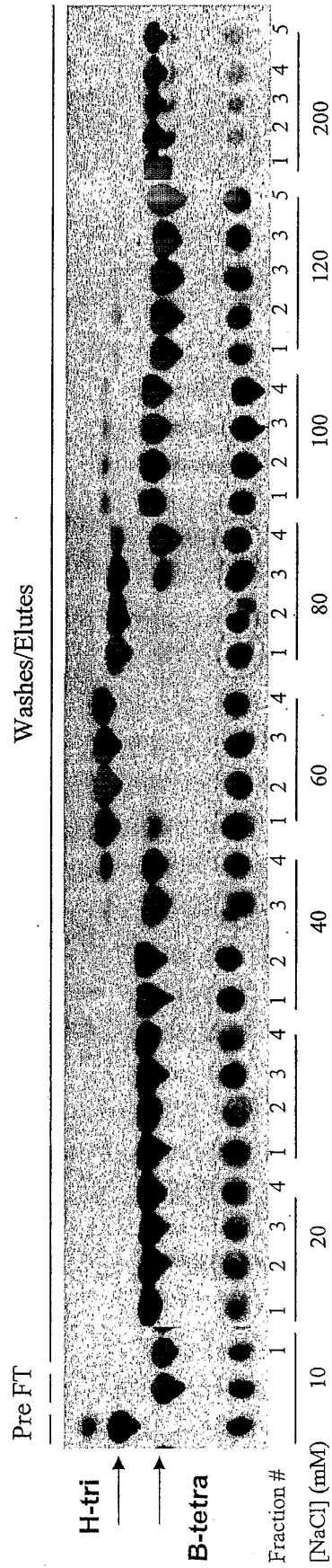
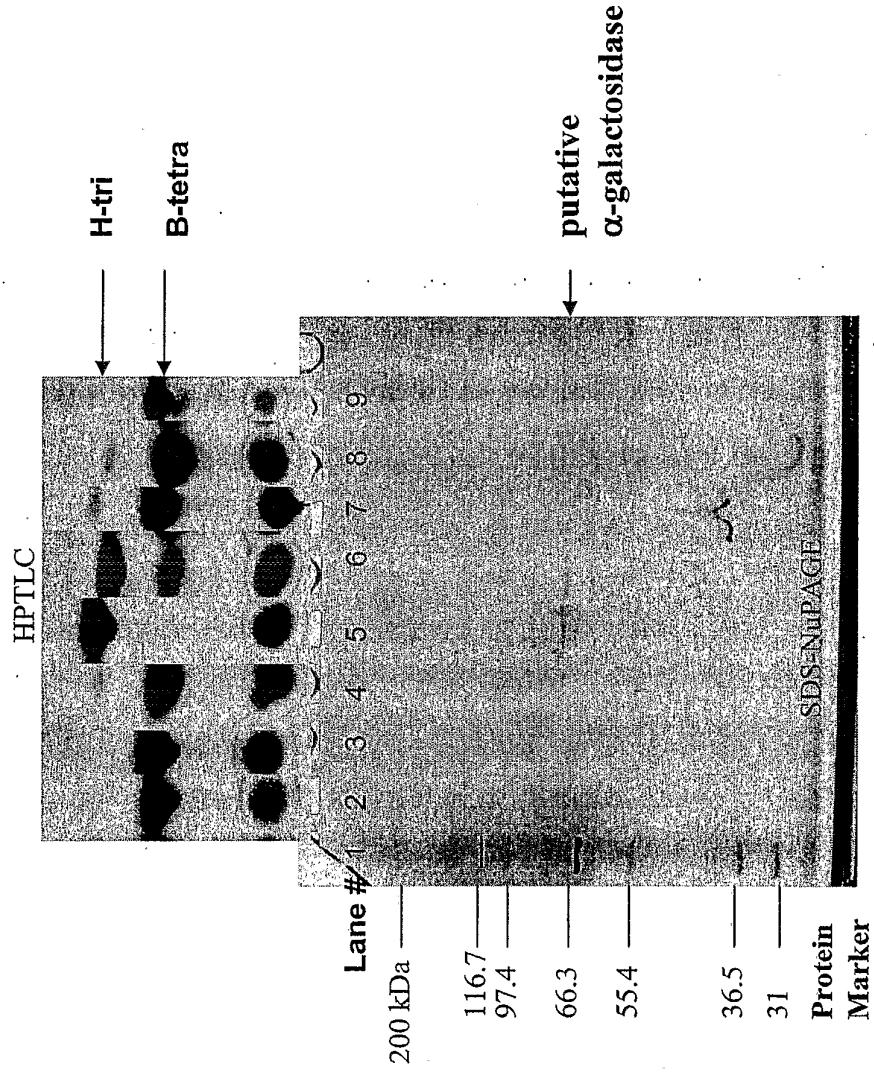


FIG. 8.



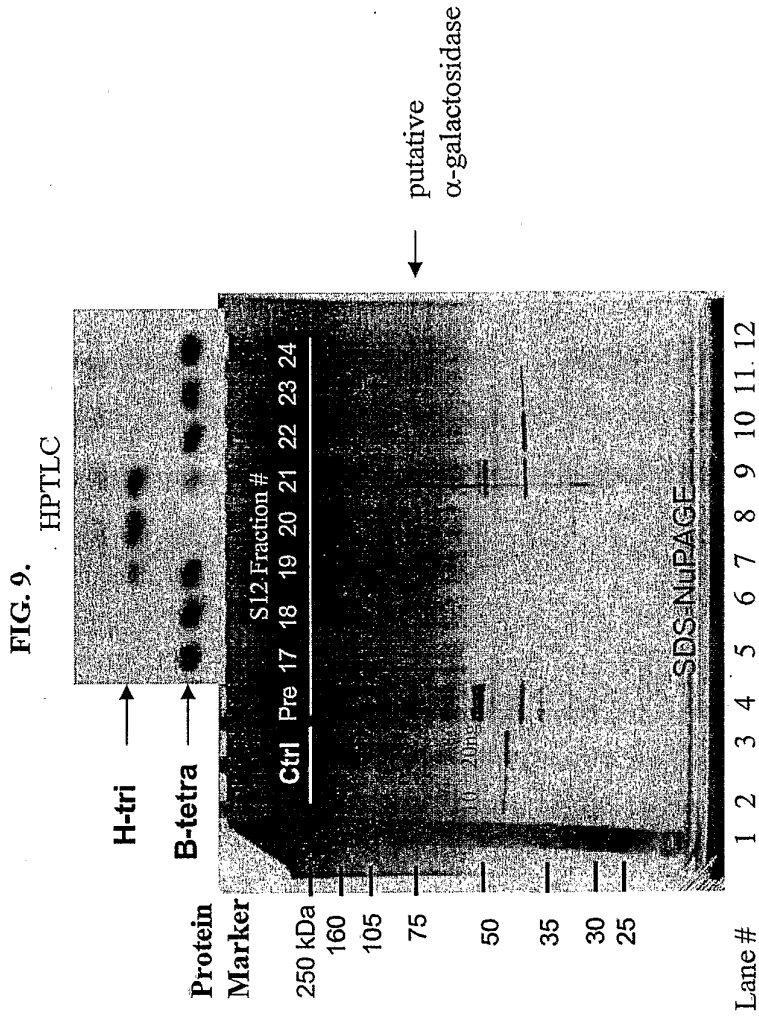


FIG.10.

1 MAHGCSGGAM SRFVFLGVAL ALLGGATSPA AAAPRVTPVV DVVDYGADP TGRtDStpAV AAALrHAKSV DRPVRIVFSK
 81 GTYQLYPERA ETRELYMSNT VGADQRYRDK KIGLLVEDMH DVTVDGGGAK LVHHGLQIAT ASIRSDVTF QNFSDYAAP
 161 EVIDATVATT GVTDGHAYRV LKIPAGSPYR VNGTHITWLG ETSPATGOPY WSGVDGLQYT QIHDPEAQRT WRGDNPLFND
 241 VAAVTDLGG RIRIDYTTAA RPADAGLVYQ MRLIERTEPG AFIWESKNVT MRSMNAYLQ SPGVVGFSE NISIDKVNFA
 321 PDRSRGRSTA SFADFVQMSG VKGKVSITRS LFDGPHDDPI NIHGTYLEV V GKPGPSTLL AYKHPQTAGF PQFAPGDEVE
 401 FATKRIMTPL ADAHAQVTAV DGPSGMDHTK PLTMTMTVTFD RFPVAGVETG GTVVENTIAT PSVVISGNVF RNVPTRGILV
 481 TTRKPVLIIG NREFDGMSTAS IYVSADAYQW YESGPPVADLT IRGNSFTRPS GPVIFVEPTN QVIDPATPVH HNISVEHNSF
 561 DIGDVTVVNA KSVGGFAFTG NTVRRLDGAD HPPYTSPLFV FHGSSGIRIA RNHYDKGLNT SVVTD

FIG. 11.

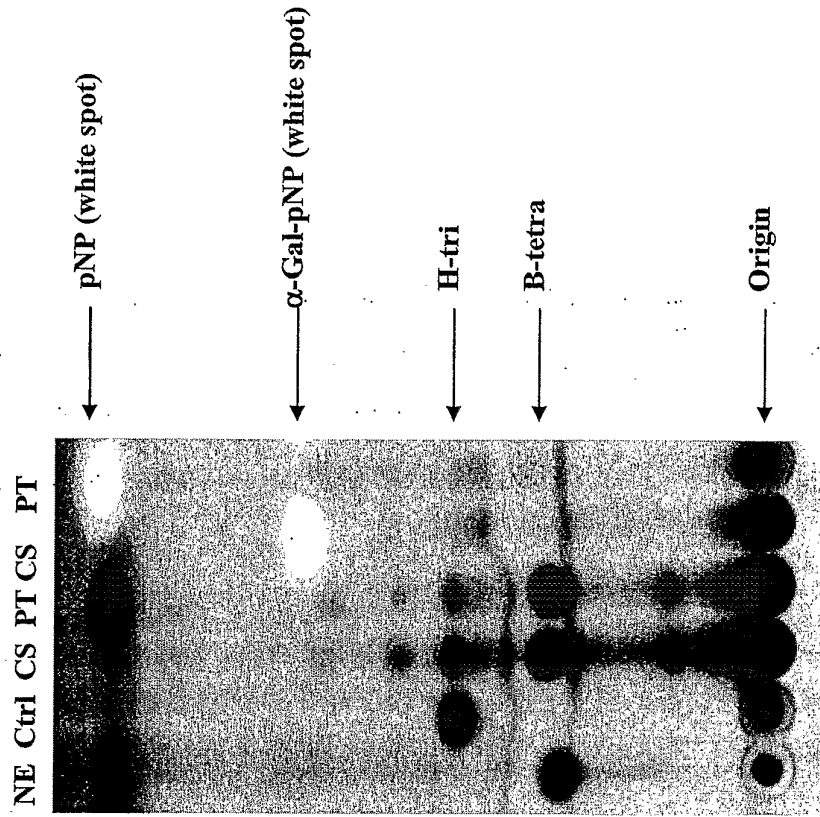


FIG.12.

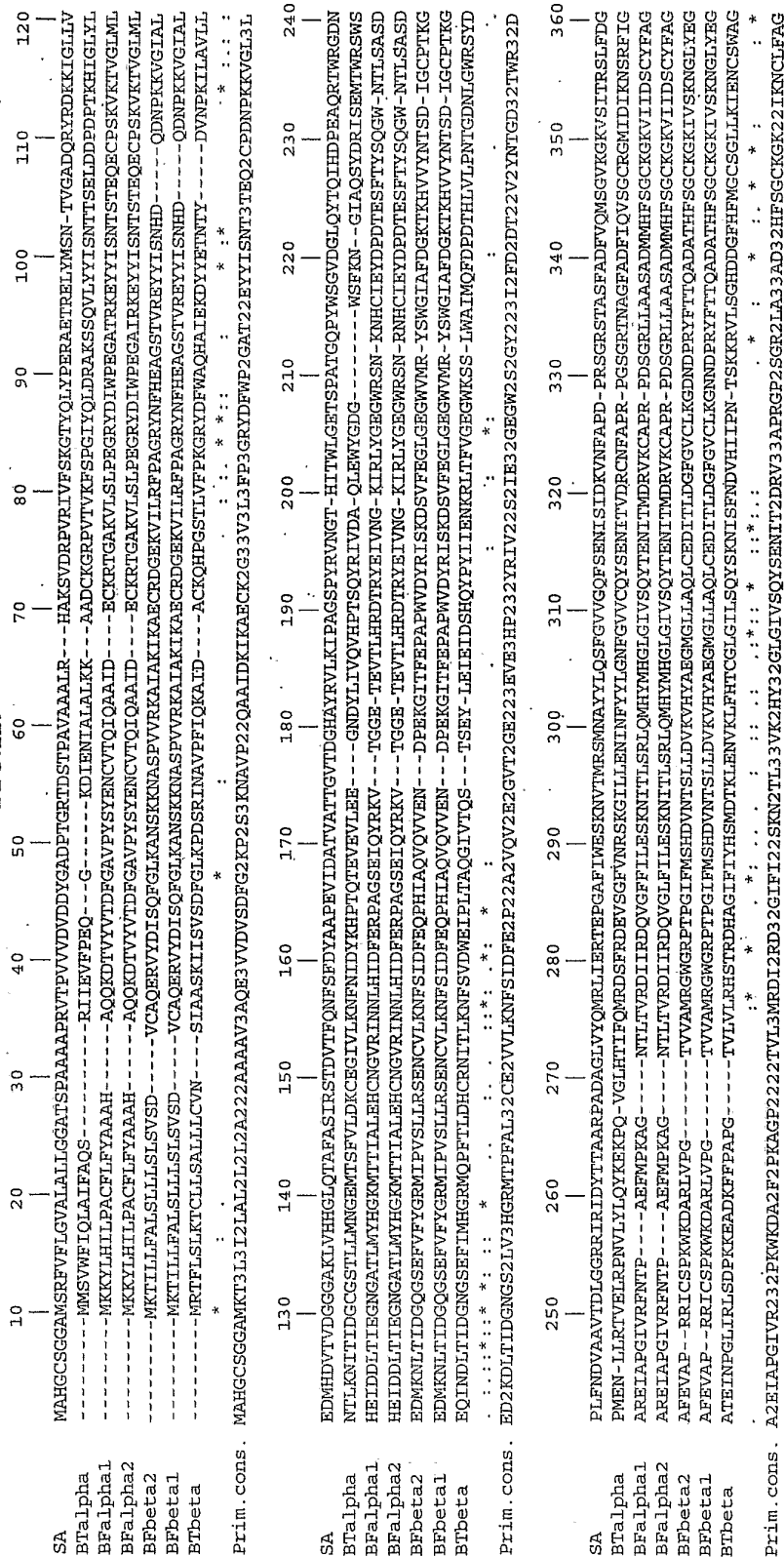


FIG. 12. continued

	370	380	390	400	410	420	430	440	450	460	470	480
SA	PHDDP	INHGTYLEVVGKPGPSTIITLAKYKHCPTAGFPQFAPGDEVEFAKRTMTPLADAHAAQTAVDGP	SGMDHFKPLTMTMTWVDRFPV-AGVETGGVTVENTATP	SVVLSGNVFRNV								
Btalpha	AHDDP	INIHGTHLRVIEFLSDNRLKIRFMHQDTFGFAFKGDDIE	LVDSRSLVVGCKVKAKLVTPEMELTILSSPLSSEVMQO	-----	KDLV	ENVTWTEVR	LINNYFARY					
Bfalpha1	AQDDP	VNVHGTMRALEKIDAQTILKRFMHGQSYGNAYFKGDTVA	FRATMERFASA-----	TVR-----	DVR	ISDRIVEVRFDRDIP-TSLELNHDCVENMTCTPEVE	LRNCYFTRT					
Bfalpha2	AQDDP	VNVHGTMRALEKIDAQTILKRFMHGQSYGNAYFKGDTVA	FRATMERFASA-----	TVR-----	DVR	ISDRIVEVRFDRDIP-TSLELNHDCVENMTCTPEVE	LRNCYFTRT					
Bfbeta2	MDDA	INVHGTYLVKVRVDDHHLI GRVMHQDWSGFEWGRPGDDVQV	FVRS	TME	LIGKQMLTAIRP--	YDKGEI	QGAREFSITFKEAIDA	PAINEKSGFGIE	MLTWTPEVLF	FAGNTRIN		
Bfbeta1	MDDA	INVHGTYLVKVRVDDHHLI GRVMHQDWSGFEWGRPGDDVQV	FVRS	TME	LIGKQMLTAIRP--	YDKGEI	QGAREFSITFKEAIDA	PAINEKSGFGIE	MLTWTPEVLF	FAGNTRIN		
Btbeta	LADDP	INIHGTCSRIMEVLSPTRIKCKFMQDSEGMEWGRPDETI	GFIEHKTRIVATG-----	KMN-----	KFEAL	NKAEFI	LSVPLP--	AGVEAGY	VIENTICT	EDAE	IRNCHFGSC	

	490	500	510	520	530	540	550	560	570	580	590	600		
SA	PTRG	LLVTRK	EVLTGNR	FDGMSASITYSADAYQWY	ESGPFVADL	ITR	SGNSFTR-----	PSGPF	IVFVPT	NQVIDP	ATPVHHN-----	ISVHN	SFDIGDVT	VVNAKSVGGFAFTG
Btalpha	PTRG	LLVTRK	EVLTGNR	FDGMSASITYSADAYQWY	ESGPFVADL	ITR	SGNSFTR-----	PSGPF	IVFVPT	NQVIDP	ATPVHHN-----	ISVHN	SFDIGDVT	VVNAKSVGGFAFTG
Bfalpha1	STRG	TLVTRPRK	VVLENNYTK	GTMSAL	LIED	AEGWY	SGPVK	DVLI	KGNT	FID--	CAYNGG	P	GHAV	IAIHP
Bfalpha2	STRG	TLVTRPRK	VVLENNYTK	GTMSAL	LIED	AEGWY	SGPVK	DVLI	KGNT	FID--	CAYNGG	P	GHAV	IAIHP
Bfbeta1	RARG	TLEST	PKT	VVED	NLDH	TSGT	ALLCGD	CNGW	FTGAC	RDV	TIR	RNF	IN--	ALTN
Btbeta	RARG	TLEST	PKT	VVED	NLDH	TSGT	ALLCGD	CNGW	FTGAC	RDV	TIR	RNF	IN--	ALTN

	610	620	630	640	650	660
SA	NVRR	L	D	G	A	D
Btalpha	NVRR	L	D	G	A	D
Bfalpha1	NLIY	S	L	D	T	E
Bfalpha2	NTIV	R	T	F	F	P
Bfbeta1	NVLK	T	N	T	F	F
Btbeta	NTL	R	D	T	T	Y

	670	680	690	700	710	720	730	740	750	760	770	780	790	800
SA	RTRG	TLVTRP	2KV	WEN	N	T	F	2	3	T	S	M	S	A
Btalpha	RTRG	TLVTRP	2KV	WEN	N	T	F	2	3	T	S	M	S	A
Bfalpha1	NTI	R	2	R	3	T	T	F	K	P	H	3	N	K
Bfalpha2	NTI	R	2	R	3	T	T	F	K	P	H	3	N	K
Bfbeta1	NTI	R	2	R	3	T	T	F	K	P	H	3	N	K
Btbeta	NTI	R	2	R	3	T	T	F	K	P	H	3	N	K

Prim. cons. AMDDPINVHGTYLAEVIEK3DD2TLKLRFMHDSQ2GFE2G2PGD2V3FVRS3TME23AKANQIT2R2P2PYDK2E32S2REFS2TFDR3IPPA22H22G2VIENLT2TPEVEIRNNVFR23

FIG. 13

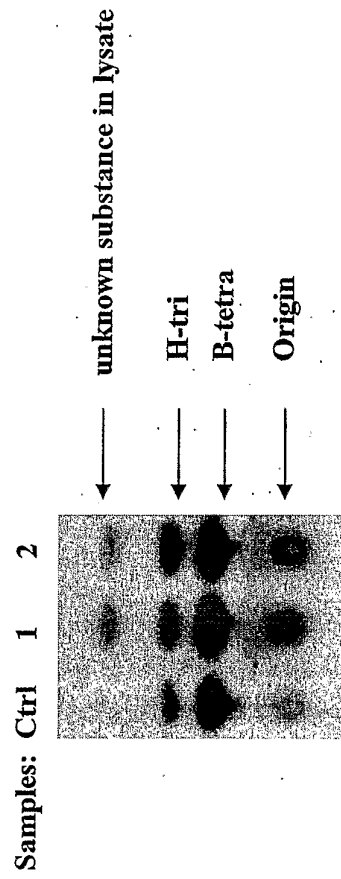


FIG. 14.

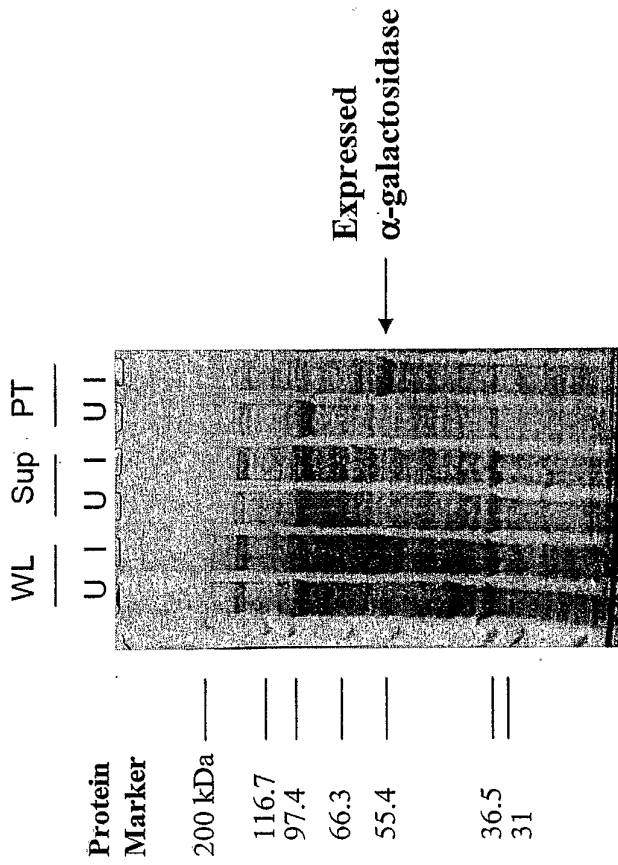


Fig. 16

```
1  MGTATAQPAL  RPQTSTVIGG  LHGAAVLDNT  GRTVIDVTFD  GADPSGKADS
51  AA AVSAAMAH  AKTVGGPTTL  HFPTGTYHIW  PERTPKRELY  VSNTVGSQDA
101 FRTKNIGILV  EDMRDVVVDG  GGSRIVNHGF  QTVFAAIRSS  DVRFTNFSQT
151 WVAPKTVDIT  VADAGVVSGQ  AYRIIDIPET  YDYAVEGTSV  RWNGERG PAT
201 GQPYWTGTNS  FDYSQVHDP  A  TNRTWR TSNP  VFPERHEDHR  PRRRQVRITY
251 GDSTAPGDRG  YVYQMREVT  R  DTPGALFWES  SRVTVDHLRL  GYLHG
301  DRGSG  RVTSGFADHI  QMSGVKGI  ITNSVEDNPQ
351  LQATAAERET  LQLRYMHNET  SGFPQFY PGD  TIELVDKRTM
401 LAAPGATAKV  VSVTGPTGSG  VPAGTDPDTY  LRTMTVVLDR  TLPAAVLAAP
451 GDYVAENTTY  TPTVEITGNT  FOAVPTRGIL  VTTRRPVRIE  NNRFDGMSMA
501 SIYISSDARS  WYESGPVRNV  TIRGNVDRP  ASPVIFFDPT  NQDFVAGO
551  EDND  FNLTGGTILS  GRGVGGLTFR  DNRVERYPHL  RLTGPSRALR
601 VGDTTTTVTTD  APPSHTSPL  FTFDGADDIT  LANNTYGN GF  NKRVTANMD
651 VSEITVTADG  LALNADSISS  APVAVSYS S  RPKVATVDSE  GVVKALSGGT
701 TSITARATIG  GVRVTSNPVK  VVVATER
```

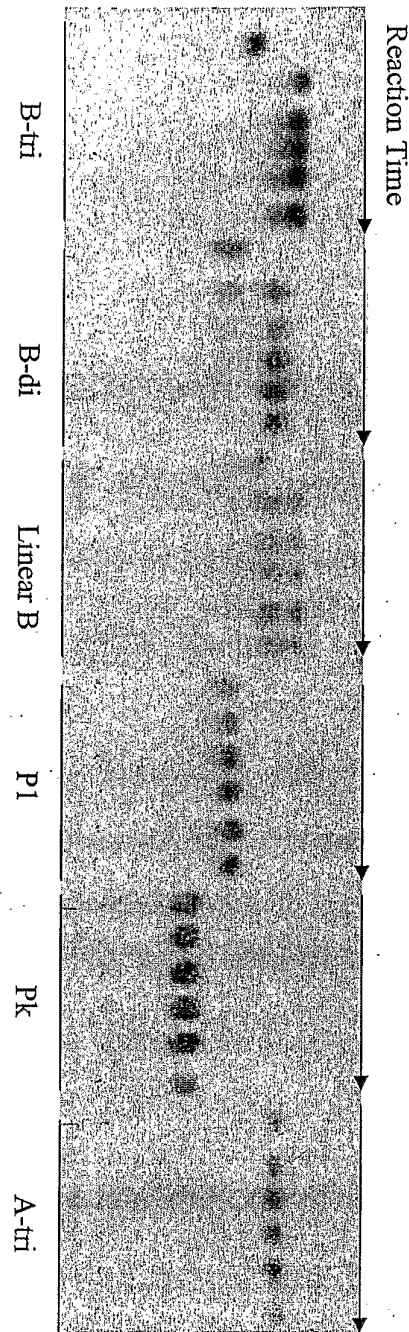


FIG.17

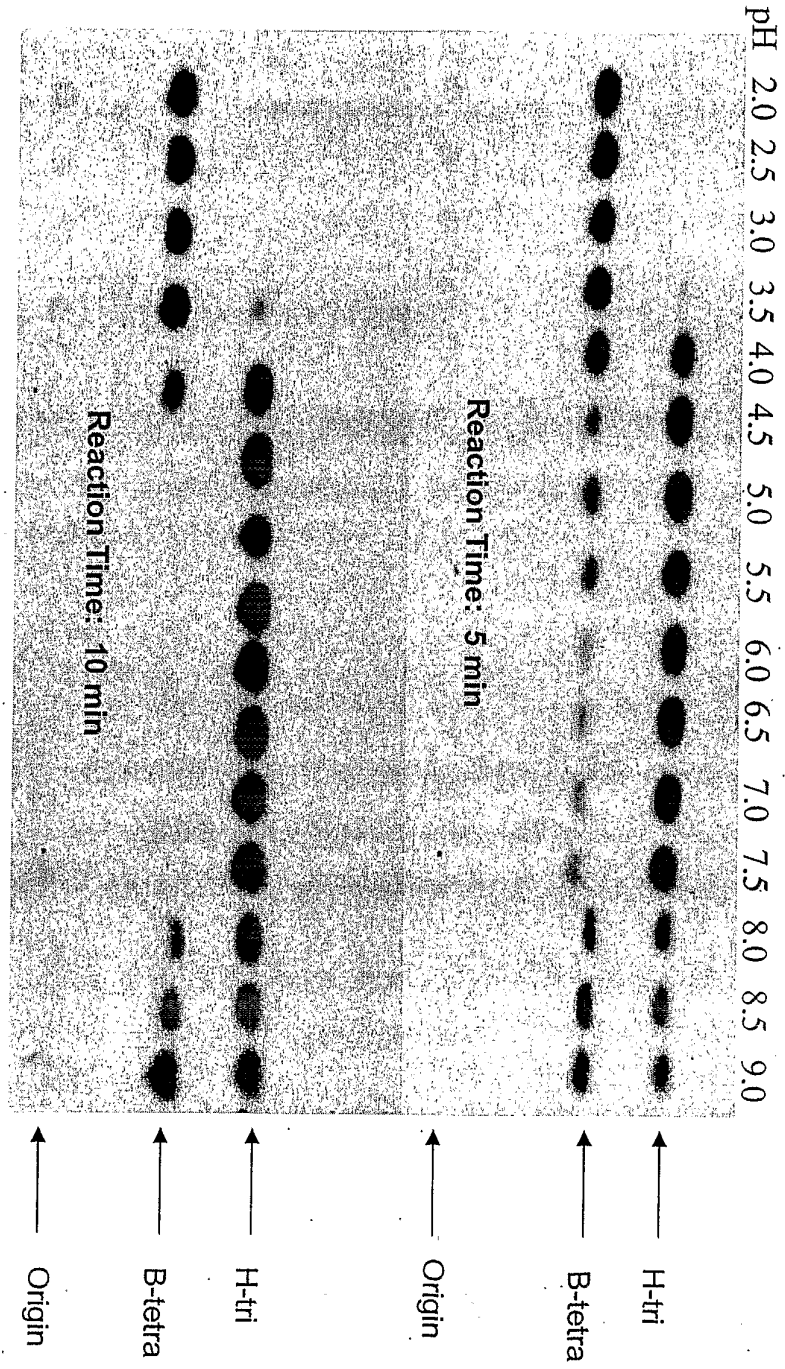


FIG. 18

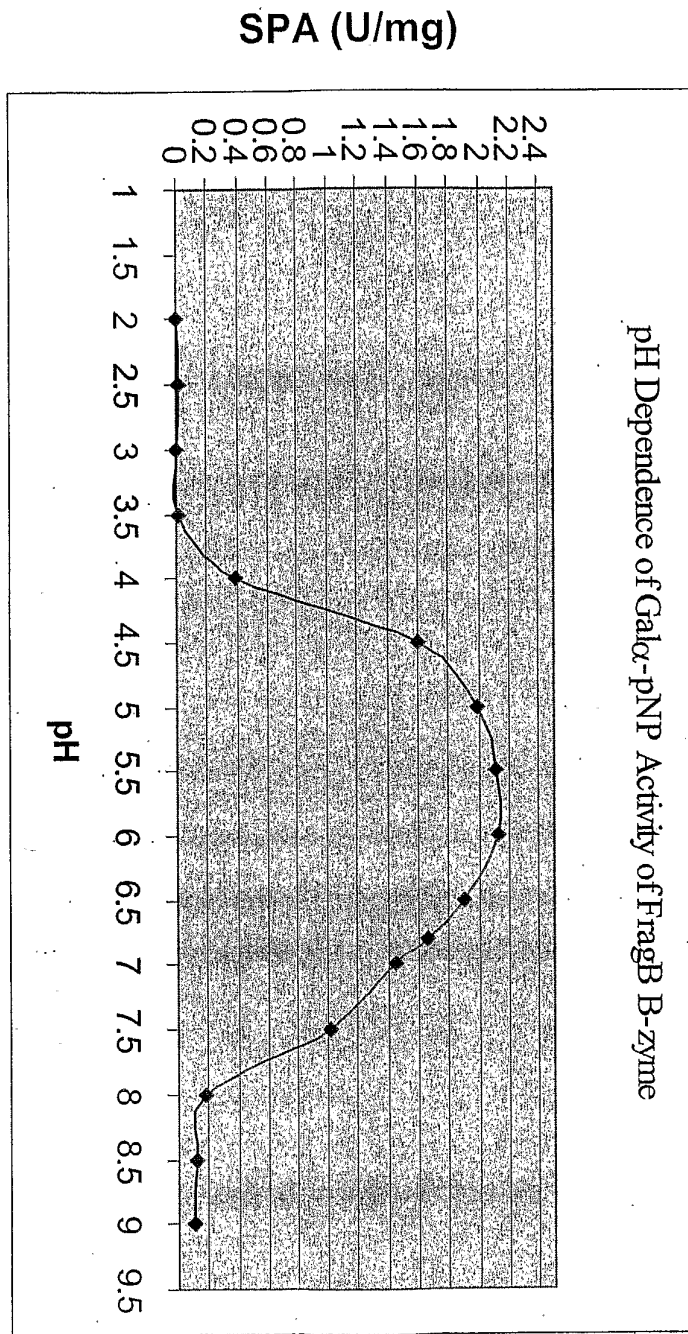
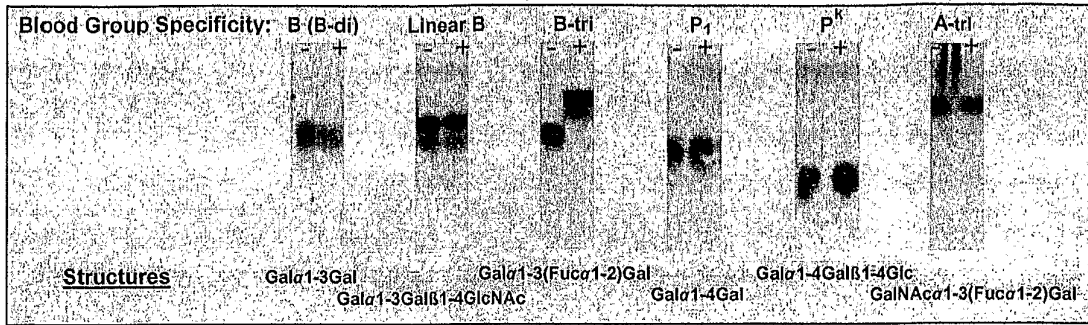


FIG. 19

Fig. 20.

A. Substrate specificity of BF α 2 (FragA) α -galactosidase.



B. Substrate specificity of BF β 1 (FragB) α -galactosidase.

